

Systematic Review

Autologous Tooth Graft: Innovative Biomaterial for Bone Regeneration. Tooth Transformer[®] and the Role of Microbiota in Regenerative Dentistry. A Systematic Review

Angelo Michele Inchingolo ^{1,†}, Assunta Patano ^{1,‡}, Chiara Di Pede ¹, Alessio Danilo Inchingolo ¹, Giulia Palmieri ¹, Elisabetta de Ruvo ¹, Merigrazia Campanelli ¹, Silvio Buongiorno ¹, Vincenzo Carpentiere ¹, Fabio Piras ¹, Vito Settanni ¹, Fabio Viapiano ¹, Denisa Hazbala ¹, Biagio Rapone ¹, Antonio Mancini ¹, Daniela Di Venere ¹, Francesco Inchingolo ^{1,*}, Maria Celeste Fatone ^{2,*}, Andrea Palermo ³, Elio Minetti ⁴, Felice Lorusso ⁵, Antonio Scarano ⁵, Salvatore Sauro ^{1,6}, Gianluca Martino Tartaglia ⁷, Ioana Roxana Bordea ⁸, Gianna Dipalma ^{1,‡} and Giuseppina Malcangi ^{1,‡}

¹ Department of Interdisciplinary Medicine, University of Bari "Aldo Moro", 70124 Bari, Italy

² PTA Trani-ASL BT, Viale Padre Pio, 76125 Trani, Italy

³ College of Medicine and Dentistry Birmingham, University of Birmingham, Birmingham B4 6BN, UK

⁴ Department of Biomedical, Surgical, and Dental Science, Università Degli Studi di Milano, 20122 Milan, Italy

⁵ Department of Innovative Technologies in Medicine and Dentistry, University of Chieti-Pescara, 66100 Chieti, Italy

⁶ Dental Biomaterials and Minimally Invasive Dentistry, Department of Dentistry, Cardenal Herrera-CEU University, CEU Universities, C/Santiago Ramón y Cajal, s/n., Alfara del Patriarca, 46115 Valencia, Spain

⁷ UOC Maxillo-Facial Surgery and Dentistry, Department of Biomedical, Surgical and Dental Sciences, School of Dentistry, Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico, University of Milan, 20100 Milan, Italy

⁸ Department of Oral Rehabilitation, Faculty of Dentistry, Iuliu Hațieganu University of Medicine and Pharmacy, 400012 Cluj-Napoca, Romania

* Correspondence: francesco.inchingolo@uniba.it (F.I.); dott.celestefatone@gmail.com (M.C.F.); Tel.: +39-331-211-1104 (F.I.); +393479914635 (M.C.F.)

† These authors contributed equally to this work as first authors.

‡ These authors contributed equally to this work as last authors.

Citation: Inchingolo, A.M.; Patano, A.; Di Pede, C.; Inchingolo, A.D.; Palmieri, G.; de Ruvo, E.; Campanelli, M.; Buongiorno, S.; Carpentiere, V.; Piras, F.; et al.

Autologous tooth graft: Innovative biomaterial for bone re-generation. Tooth Transformer[®] and the role of microbiota in regenerative dentistry. A Systematic Review. *J. Funct. Biomater.* **2023**, *14*, 132. <https://doi.org/10.3390/jfb14030132>

Academic Editor: John W. Nicholson

Received: 14 January 2023

Revised: 16 February 2023

Accepted: 25 February 2023

Published: 27 February 2023



Copyright: © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Different biomaterials, from synthetic products to autologous or heterologous grafts, have been suggested for the preservation and regeneration of bone. The aim of this study is to evaluate the effectiveness of autologous tooth as a grafting material and examine the properties of this material and its interactions with bone metabolism. PubMed, Scopus, Cochrane Library, and Web of Science were searched to find articles addressing our topic published from 1 January 2012 up to 22 November 2022, and a total of 1516 studies were identified. Eighteen papers in all were considered in this review for qualitative analysis. Demineralized dentin can be used as a graft material, since it shows high cell compatibility and promotes rapid bone regeneration by striking an ideal balance between bone resorption and production; it also has several benefits, such as quick recovery times, high-quality newly formed bone, low costs, no risk of disease transmission, the ability to be performed as an outpatient procedure, and no donor-related postoperative complications. Demineralization is a crucial step in the tooth treatment process, which includes cleaning, grinding, and demineralization. Since the presence of hydroxyapatite crystals prevents the release of growth factors, demineralization is essential for effective regenerative surgery. Even though the relationship between the bone system and dysbiosis has not yet been fully explored, this study highlights an association between bone and gut microbes. The creation of additional scientific studies to build upon and enhance the findings of this study should be a future objective of scientific research.

Keywords: tooth transformer[®]; biomaterials; autologous graft; tooth graft; demineralized dentine matrix; microbiota; osteogenic material; regenerative dentistry; bone remodeling; morphogenetic proteins

1. Introduction

Regenerative medicine has received a lot of attention and development in the medical field in recent decades. The study of biomaterials has yielded excellent and predictable results, thanks partly to biotechnology.

The study of bone tissue regeneration, which is widely used in dentistry, has piqued the researchers' interest [1]. Graft material must function as an osteoconductive scaffold, provide a mineral substrate, contain osteoinductive cells, platelet growth factors, and cell differentiation factors, and be decontaminated [2,3].

Furthermore, the grafts must not only fill and occupy the space of the bone defect, but also maintain the clot and/or provide proteins and/or release cells contained within them. In order to allow the adhesion of osteoblasts and osteoclasts and favor osteoid formation in the first phases of bone regeneration, the granules must have a specific size (500–1000 μm), be manageable and plastic during the surgical phase, as well as have spaces between them that can allow rapid neovascularization [4].

Based on their properties and origins, scaffolds are classified into [5]:

- autologous or allographic human bone, i.e., taken from another donor (4%)
- xenographic: animal bone, coral (54%)
- alloplastic: bioglass, hydroxyapatite (HA), tricalcium β -phosphate (TCP), polyglycolic, and polylactic acid derivatives (33%)
- other (9%)

All studies have confirmed that autologous bone grafting is the gold standard.

This type of graft cannot induce adverse immune reactions because it is autologous, has osteogenic, osteoinductive, and osteoconductive capabilities and stimulates osteoproliferation with rapid healing. However, the need for an additional invasive operation, increased morbidity, pain, and the difficulty of the harvesting areas, reduce the rates of use of autologous bone grafting [6]. Furthermore, growth factors are released only after the graft has been completely resorbed [7,8].

Therefore, grafts of various origins (allographic, xenographic, and alloplastic) have been used for regenerative techniques [9,10]. The majority of these grafts only have osteoconductive properties and very long resorption times (coral and HA) or short resorption times (polyglycolic and polylactic acid derivatives), while others pose risks of immune reactions and infections (allografts), are incompatible with religious cultures, and are expensive on the market, despite all studies showing the good bone regenerative efficacy of these grafts [11,12].

In recent years, special attention has been given to dentine as a biomaterial in bone regeneration [13]. An analysis of its biochemical composition has revealed similarities with bone tissue. Dentine, like bone tissue, is 61% inorganic ($\text{HACa}_5(\text{PO}_4)_3\text{OH}$), with crystals that are 10 times larger than bone and 300 times smaller than enamel. A total of 90% of the organic component (39%) is collagen: collagen type I (95%), and collagen types III, V, and XII (5%). Collagen gives resilience and elasticity and makes the structure resistant to fractures [14].

The 10% consists of non-collagenous proteins (Osteopontin OPN, Dentin sialoprotein DSP, DGP, DPP–Bone sialoprotein (BSP), Osteocalcin, Dentin sialoprotein DSP, Dentin matrix protein-1 DMP-1, Collagen type 1, Cbfa1 RUNx2, Bone morphogenetic protein BMP-2, and Transforming growth factor β (TGF- β) 19). Most of the proteins contained in the dentine are also present in bone [15–18].

One of the earliest studies on the tooth as a biomaterial for grafting is that of Schmidt-Schultz and colleagues, in which using teeth from the late pre-ceramic Neolithic period (approx. 8000 years ago), teeth from the early Middle Ages, and recently extracted teeth, aided in the identification and isolation of growth factors such as Insulin Growth Factor-II (IGF-II), Bone Morphogenetic Protein-2 (BMP-2), and TGF- β [19].

The proteins, quantified and isolated by electrophoresis, were still present undenatured and therefore active even after 8000 years, and the quantity did not differ between the three groups. The results of this study, therefore, reported that the proteins of the

extra-cellular matrix of bone and tooth are protected from the aggression of physical and chemical agents even after death by HA, and their osteoinductive capacities can be utilized after resorption of the mineralized part following the regenerative process [20,21].

Thirteen types of BMP proteins are recognized. They can activate the process of bone neoformation even in heterotopic locations [22]. Specifically, BMP-2 induces differentiation into the osteoblasts of mesenchymal cells. BMP-3 stimulate bone formation, BMP-7 stimulate differentiation into osteoblasts [23]. BMPs initially activate the replication and migration of mesenchymal cells from surrounding tissues to the regenerated area and then repress the WNT signal that blocks the differentiation of mesenchymal cells [5] (Figure 1).

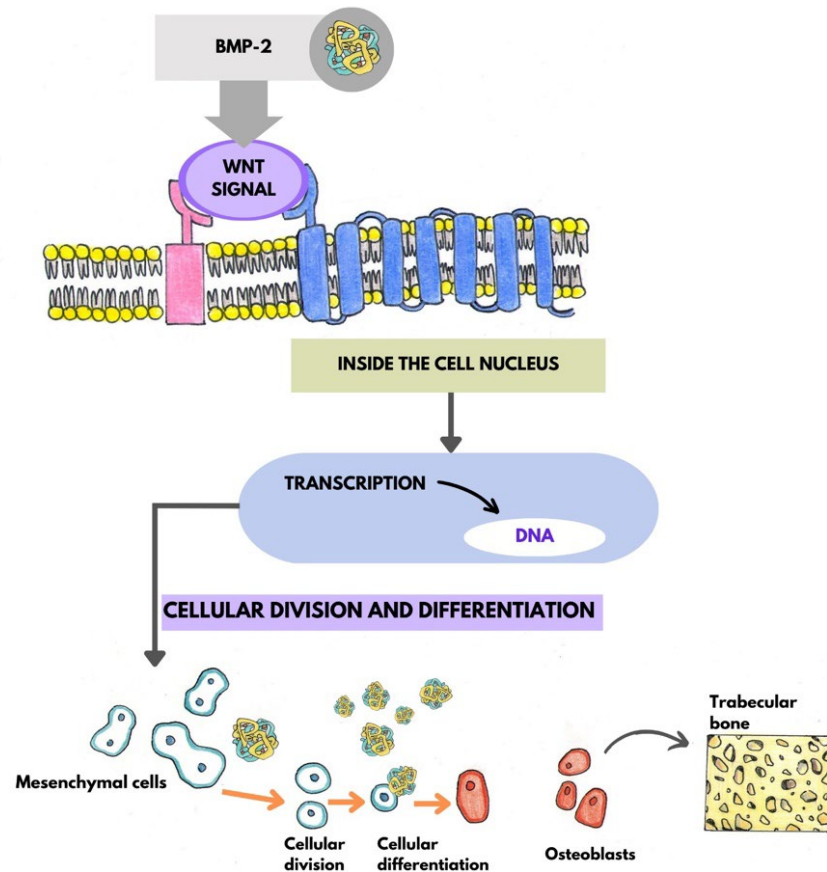


Figure 1. Mechanism of BMP-2, cellular division and differentiation into osteoblasts.

BMP-2 binds to the cell membrane via the Bone Morphogenetic Protein Receptor Type 1 (BMPR1A), a transmembrane protein of the serine/threonine kinase group, and activates the transformation of mesenchymal cells into osteoblasts that initiate the neo-attachment of bone. BMPs are soluble proteins and require an insoluble carrier, collagen type 1, to activate mesenchymal cells. Individually, the two proteins do not stimulate bone formation [23,24].

IGF-1 and IGF-2 are growth factors activated by Growth Hormone (GH) and are incorporated into the bone matrix. They stimulate cell proliferation and their functions, such as that of collagen type I production. During resorption, they are released, activating osteoblast production and the remodeling of new bone [5,25].

Non-collagenous proteins, which make up 10% of the proteins in dentin, are growth factors that are not specific to the bone but are necessary for its neoformation [26]. OPN controls bone homeostasis, and binding to HA ensures the mineralized base of the bone. BSP stimulates angiogenesis ensuring the neovascularisation essential for the apposition

of new bone. Osteocalcin, which indicates bone metabolism, intervenes by activating bone metabolism. A Cbfa Run X2 intervenes in osteoblast differentiation [27–30].

The mechanisms of stimulation in bone regeneration promoted by demineralized dentin are quite similar to the formation of new bone by autologous bone. After demineralization, both demineralized bone matrix (DBM) and demineralized dentin matrix (DDM) contain type I collagen, growth factors, and bone morphogenetic proteins (BMP-2) [31].

Different techniques for demineralizing teeth were analyzed with equally different results in the production of bone tissue [32]. The degree of sterilization, the repeatability of the system, the liquids and their concentration, the degree of demineralization, the size of the granules, the amount of residual protein after treatment, the wettability and plasticity of the granules, and the ergonomics of the system must be considered when evaluating the various tooth crushing systems [4].

The Tooth Transformer (TT[®]) device proved to be the best-performing system. The need to first prepare the tooth by hand (cleaning and reduction into fragments by milling) and then place it in the grinding container remains a drawback [4]. Intact teeth, devitalized teeth, and even deciduous elements were considered.

Data reported that osteoblasts only adhered to the demineralized surface. Different levels of demineralization are expressed in the Ca/P ratio. The ideal Ca/P ratio in human bone is 1.67 [33–35]. A granule size between 400 and 800 μm (constant size in TT[®]) would be the most efficient for the quality and speed of the bone produced (4–6 months) [35,36]. Partially demineralized dentin serves both as a supporting filler and maintains the osteogenic potential of proteins, BMPs, and collagen [37–39].

The sterilization of the product and the maneuverability of the six solutions used automatically in sequence by the TT[®] elevate its quality. The size of the bone defect and the small amount of autologous dental graft available could be a limitation to the use of the system. In this case, either autologous grafts (platelet derivatives together with autologous CGF membranes) and/or xenografts (bioss) or alloplastic grafts (HA or TCP) have been used, exploiting both osteoinductive properties (platelet derivatives and GF) and the scaffold function of the others [39–42].

The presence of dysmetabolic diseases, such as diabetes, and the increasing age of patients, who often have comorbidities and undergo implant-prosthetic therapies, could affect the bone regenerative response [43]. Recent studies associated the human microbiota with the proper functioning of the entire metabolism and thus a state of well-being. The microbiota refers to several microorganisms larger than the number of cells in the human organism living in symbiosis. They are largely present in the gut (95%), in other parts of the digestive system, in the genital system, and in the eyes and ears, skin, etc. [43–46].

The genetic composition of the microbiota is called the microbiome [47]. The intestinal microbiota, with its metabolites (short-chain fatty acids SCFA and intestinal wall lipids) interacts with resveratrol (RSV), a naturally-occurring polyphenol, whose anti-inflammatory, anti-cancer, antioxidant and immunostimulant properties improve the clinical consequences of metabolic syndrome, which is frequent in industrialized and developed populations, characterized by obesity, hypertension, hyperglycemia, and cardiovascular disorders [48–52].

The microbiota, with its barrier effect on the intestinal epithelium, reduces the production of inflammatory cytokines and influences the immune system by reducing osteoclastic activity [53,54]. The gut microbiota is also attributed to the ability to produce the hormone IGF-1, which regulates osteoblast differentiation enhanced by the intake of pre- and probiotics [55–57]. The microbiota modulates the absorption of vitamins such as folic acid and the vitamin B2–B12 complex. It synthesizes vitamin K, a fundamental carrier of vitamin D [58–60]. Vitamin D3 activates TGF β -1 and TGF β -2, which are contained in the bone matrix, and which in turn enhance the activation of BMP-2 with an osteoinductive effect [22].

Primary osteoporosis, a major bone disease, appears to be related to the gut microbiota via the gut–brain axis, with mechanisms that are still unclear [61]. In particular, there is an increase in the Dialister and Faecalibacterium genera in the gut microbiota of patients

with primary osteoporosis [62]. A valid diagnostic and/or therapeutic guideline in clinical practice could be necessary to monitor the composition of the gut microbiota [63].

The purpose of this study is to evaluate the efficacy of autologous tooth as a grafting material and to investigate its properties and interactions with bone metabolism.

2. Materials and Methods

2.1. Search Processing

This systematic review has been performed in accordance with the principles of the PRISMA and International Prospective Register of Systematic Review Registry guidelines (ID 390491) [64]. The literature search was performed on PubMed, Cochrane Library, Scopus and Web of Science databases from 1 January 2012 up to 22 November 2022, with English-language restriction. A combination of words that matched the purpose of our investigation, whose primary focus is the use of grafts with autologous materials of dental origin was used; hence, the following Boolean keywords were chosen: (“demineralized dentin matrix” OR “tooth graft”) AND “biomaterial”.

2.2. Inclusion Criteria

Reviewers worked in duplicate and analyzed all suitable trials, considering the following inclusion criteria: (1) studies only on humans; (2) open-access studies; and (3) studies that analyzed the use of grafts with autologous materials of dental origin, including articles in which this is associated with other materials and possible associations with systemic diseases, microbiota or intake of RSV, curcumin and quercetin. Articles dealing with dental grafts using non-autologous materials were excluded, as were articles in non-English languages.

3. Results

A total of 1516 articles were identified from Pubmed (755), Scopus (537), Cochrane Library (85), and Web of Science (139) databases, which led to 1168 works after removing duplicates (348). Nineteen relevant publications were added by searching the reference list of eligible papers. A total of 1117 articles were excluded by analysis of the title and abstract. The remaining 51 articles were added to the 19 papers found by reference list, leading to 70 publications that were assessed for eligibility by the authors. A total of 33 publications were excluded because they were off-topic. A final 18 studies were included for qualitative analysis of the review (Figure 2) (Table 1).

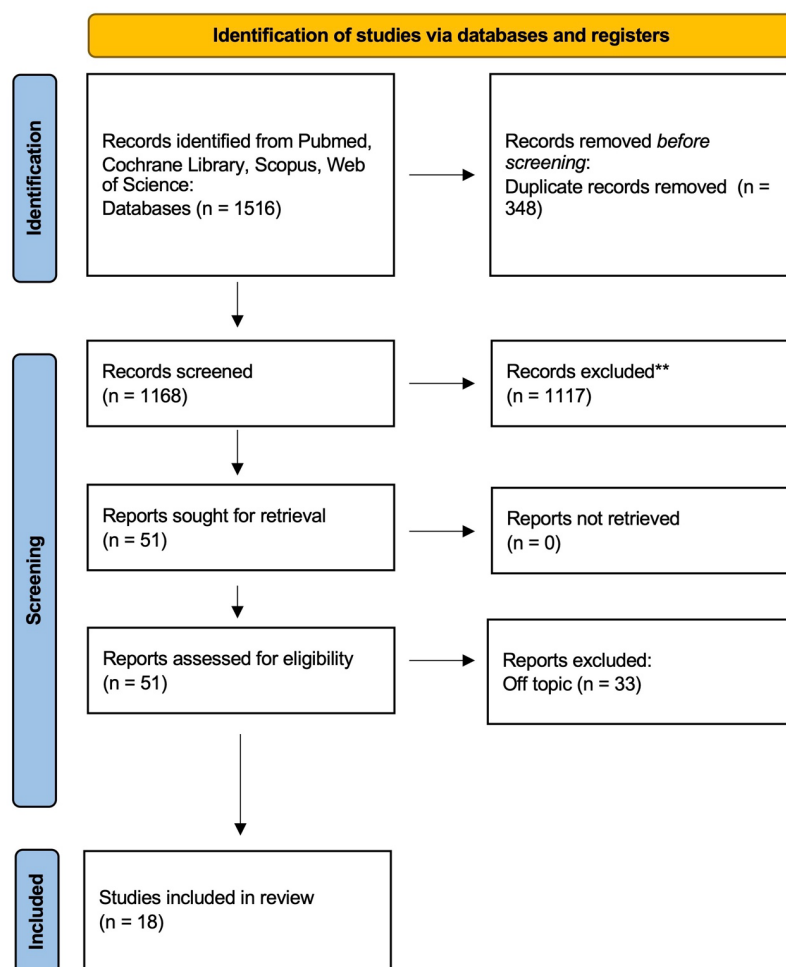


Figure 2. PRISMA flowchart diagram of the inclusion process.

Table 1. Descriptive summary of item selection.

Authors (Year)	Type of the Study	Aim of the Study	Materials	Results
Tazaki et al. (2012) [65]	Clinical report	The use of autologous dentin grafts for the treatment of bone defects.	The extracted tooth was crushed by a newly developed automatic mill. The crushed granules were demineralised in 2% HNO ₃ . The granules were washed in distilled cold water and freeze-dried (size: 0.5–2.0 mm).	The human dentin can be used as an autogenous biomaterial for local bone engineering.
Kabir et al. (2014) [31]	Clinical report	These studies imply that dentin may actually replace bone as a viable biomaterial.	Case 1: a 29-year-old male patient. A #38 was used to create tooth-derived granules, which were then demineralized in 2% HNO ₃ for 30 min and thoroughly cleaned. DDM was transplanted into the bone gap. Case 2: A 20-year-old woman who had an impacted third molar (#48). The DDM autograft and extraction of the affected tooth were completed concurrently.	90%–95% of patient-own recycled dentin matrix had remodeled into bone, resulting in excellent bone defect repair.
Jun et al. (2014) [66]	A prospective clinical study	Evaluation of DDM and Bio-Oss as sinus bone grafts in patients with residual bone height less than 5.0 mm in the maxillary posterior area.	43 patients, 21 in the control (Bio-Oss) and 22 in the test group (DDM 0.5–1.0 mm). After four months the sites were analyzed with microcomputed tomography analysis and histomorphometric analysis.	In the two groups, there was no difference in bone density and height, instead, there was a significant difference in osteoid thickness.

Kim et al. (2015) [67]	Case reports	To evaluate the clinical use of the chairside-prepared demineralised tooth immediately after extraction for alveolus preservation.	The use of the extracted tooth as graft material radiographic and histological evaluation of the graft site.	The use of the dental block is effective in maintaining height and thickness of the bone in the preservation of the alveolus.
Bono et al. (2017) [68]	In vitro study	To examine the effects of demineralization on the physical-chemical and biological behavior of D and E.	Human dentin and enamel were minced into particles ($\varnothing < 1$ mm), demineralized, and sterilized. Thorough physical-chemical and biochemical characterizations of native and demineralized materials were performed by SEM and EDS analysis and ELISA kits to determine mineral, collagen type I, and BMP-2 contents. In addition, MG63 and SAOS-2 cells were seeded on tooth-derived materials and Bio-Oss®, and a comparison of cell responses in terms of adhesion and proliferation was carried out.	The demineralization process determined an increase in BMP-2 bioavailability, favouring the development of more effective, osteoinductive tooth-derived materials for bone regeneration and replacement.
Li et al. (2018) [69]	A prospective clinical study	Evaluation of DDM and Bio-Oss as bone substitutes in GBR for immediate placement of implants in periodontal post-extraction sites.	For 40 patients, DDM (size 0.5–1.0 mm) and Bio-Oss were used as bone grafts in the test group and control group, respectively. The implant stability, evaluated by Osstell Mentor, and marginal bone resorption, evaluated with x-ray examinations, were measured at T0, at 6 and 18 months after surgery.	The values of implant stability and marginal bone resorption were superimposable in the two groups.
Minetti et al. (2019) [38]	Multicenter Clinical Study	Studied the use of extracted tooth as autologous tooth graft after endodontic root canal therapies used for socket preservation and evaluated the implant insertion in regenerated bone with six-month follow up.	A total of 98 patients (29 men and 69 women) with an average age 53.7 years. Autologous tooth as a graft after the TT® device procedure.	The success rate of the tooth graft procedure was 99.1% (one site was infected and lost the regeneration and the implant). In all cases, after all implants were inserted, complete osseointegration after proper healing period was achieved. After the healing period, hard and soft tissues were stable. The healing of soft tissues after grafting procedures was free of complications. The implant success rate was 98.94% (one implant failed).
Cardaropoli et al. (2019) [70]	Case report	To evaluate the regenerative potential of particles obtained from a crushed extracted tooth.	After tooth removal, the clean root was ground, and the dentin and cementum granules were grafted into a fresh extraction socket for a ridge preservation procedure.	Tissue healing was evaluated by histologic and radiologic analysis. The volume of the ridge was preserved. Histologically, a dentin-bone complex was reported. New bone formation was evident, with an intimate contact between bone and both dentin/cementum.
Kadkhodazadeh et al. (2020) [71]	In vitro study	To evaluate the osteopromoting ability of human tooth powder and compare it with a bovine xenograft, a synthetic material, and a demineralized freeze-dried bone allograft (DFDBA).	A total of 30 teeth were collected. The samples were ground to a powder with particles $< 500 \mu\text{m}$. Osteoblast-like cells of MG-63 were cultured with vine xenograft, the synthetic the tooth powder, Cerabone, DFDBA, and Osteon graft, and the DFDBA. However, II. Cell proliferation was assessed by the MTT assay at 24 and 72 h intervals.	Tooth powder was able to increase osteogenic cell proliferation in comparison with the bovine xenograft, the synthetic its osteopromoting ability was less than the osteogenic materials.
Minetti et al. (2020) [37]	Multicenter pilot study	Evaluation of post-extraction site preservation with DDM from vital or endodontically treated teeth and a collagen membrane.	A total of 28 patients and 32 extractions. After 4 months, 32 biopsies were performed, and histological and histomorphometric analyses were evaluated.	There was no significant difference in the bone regeneration of sites where DDM was obtained from vital or endodontically treated teeth.
Minetti et al. (2020) [72]	Case reports	Present a case of alveolar socket preservation by using tooth graft material and	One 26-year-old women, nonsmoker, ASA-1 Autologous Deciduous Tooth-Derived Material.	Good results in terms of clinical and radiographic outcomes showing the absence of bone

		one implant-supported re-habilitation		resorption process and the stability of soft tissues after two years.
Sánchez-Labrador et al. (2020) [73]	A split-mouth clinical trial	Evaluation of post-extraction site preservation of the included third molar with and without DDM.	A total of 15 patients, 30 lower third molars were extracted, 15 sites post-extraction were grafted with DDM (300–1200 µm) and 15 contralateral sites were left to heal without graft. At three and six months, the sites were evaluated with x-ray and probing of periodontal tissue.	Bone regeneration and reduced periodontal defect on the distal side of the lower second molar were found in the sites grafted with DDM.
Korsch et al. (2021) [74]	Retrospective study	Using autogenous dentin for lateral ridge augmentation.	For the tooth-shell method (TST): 28 patients (15 females, 13 males) with 34 areas and 38 implants, autogenous dentin slices were taken from teeth and utilized to restore lateral ridge deficiencies. The control was the bone-shell technique (BST), which was performed on the autogenous bone on 31 patients (16 females and 15 males) with 32 areas and 41 implants. In both situations, implants were put in at the same time. A follow-up three months following implantation.	Between the two groups, there were no appreciable variations in the overall number of problems. One implant with TST and one with BST both showed horizontal hard tissue loss of 1 mm and 0.5 mm, respectively.
Cervera-Maillo et al. (2021) [75]	Prospective clinical trial	To evaluate the efficacy of extracted teeth processed into bacteria-free particulate dentin in a Smart dentin grinder and then grafted immediately into alveolus post extraction or into bone deficiencies.	Ten healthy, partially edentulous patients with few teeth in the mandible were recruited in the study. After their own teeth were grinded, particulate teeth were placed in empty sockets and bone defects after teeth extractions. Furthermore, after 3,6, 12, and 24 months, core samples using a 3 mm trephine were obtained.	Particulate dentin grafts should be considered as an alternative material for sockets' preservation, split technique, and sinus lifting. Clinically and histologically, the performance of the dentin graft is at least comparable to extensively used xenogeneic or allogenic biomaterials.
Minetti et al. (2022) [76]	Histological specimen study	The aim of the study was to explore the histomorphometric outcomes of tooth derivative materials used as bone substitute material in socket preservation procedure.	The use of a demineralised tooth as grafting material was evaluated for the preservation of the alveolus.	No significant difference was noted between the maxillary and mandibular sites, including defect type and section position.
Okubo et al. (2022) [77]	Case report	To evaluate the effectiveness of DDM for GBR in a patient with severe bone defect on the anterior upper jaw.	Ridge augmentation with DDM in a patient with atrophic maxilla and implant placement. After one year from the surgery a biopsy was executed.	Histological findings revealed the direct formation of new bone on DDM residue. Radiographs showed in the region of upper lateral incisor an increase in horizontal breadth of 3,16 mm after implant placement. The upper level's breadth grew from 3.38 mm to 5.92 mm.
Pohl et al. (2022) [78]	Case report	Assess concerns about the biological response of these ATDGs in preparation for implant placement and subsequent osseointegration.	After 12 weeks of extraction socket healing, an implant with an acid-etched surface was placed using osseodensification osteotomy preparation and was retrieved after 16 weeks of integration. Histologic analysis revealed ≥ 64% of direct bone-to-implant contact at multiple regions of interest along the implant surface. Residual dentin particles have been searched in contact with the implant.	Residual dentin particles were scarce and were never found in contact with the implant. The autologous tooth-derived grafts did not interfere with implant osseointegration.
Minetti et al. (2022) [79]	Clinical trial	Evaluation of alveolar ridge augmentation with and without collagen membrane associated with DDM.	Six patients with defects requiring bone augmentation and DDM (406 µm and 815 µm with peaks up to 1110 µm). In Group 1, DDM was associated with a resorbable membrane, and in group 2 only DDM was used. At four months, a bone tissue biopsy was performed.	Histological analysis showed more bone volume and vital bone in the sites where membrane had been used in association with DDB.

4. The Autologous Tooth Graft

Autologous bone grafts have been the gold standard for bone defect regeneration for more than a century, having an important impact on regenerative medicine. Despite the

diversity of structure between dentin and cortical bone, the biochemical composition presents similarities. The majority of dentin is composed of proteins common to both dentine and bone. Collagen guarantees resilience and elasticity, making them resistant to fractures. The 10% is made in Non-Collagenous Protein [15,16,80].

Bone induction is a biological process based on the recruitment of undifferentiated and pluripotent native cells which develop into the bone-forming cell pathway. The stimulation of new bone formation by bone autograft is quite similar to the bone regeneration favored by demineralized dentine [5]. These dynamics consist of a complex mechanism of cellular differentiation induced by the interaction of inducing cells and responding cells that stimulate new bone formation. Microscopically, dentin is reabsorbed thanks to the increase in the number of cell mesenchymal, macrophages, and osteoclasts carrying collagenolytic enzymes [5].

It has been demonstrated that the use of demineralized dentin induces invasion of the dentinal tubules by the processes of osteocytes and their cytoplasm [81]. The demineralization process of dentin increases BMP-2 bioavailability, leading to a new osteoinductive bone. In addition, the demineralization process in enamel, which is even less effective than in dentin, gives an increase in BMP-2 bioavailability and improved performance *in vitro* compared with native enamel. In this study, human dentin and enamel were reduced in demineralized and sterilized particles with a diameter of 1000 μm , [81]. Bono et al. demonstrated that sterilization is a fundamental process for grafts, even more so than the other chemical and physical treatments, and demineralization improves the osteoinductive properties of dentin [68,82].

A radiographic one-year follow-up evaluation of two case reports showed an increase in radiopacity and density on the site where a demineralized dentin graft had been used, resulting in complete restoration and healing via new bone formation without any clinical complication for the patient [31]. In another case report, a 1-year intraoral radiological evaluation enabled to appreciate stability of marginal bone levels where a conical-shaped implant has been positioned 24 weeks after tooth extraction filling the socket with dentin graft. CBCT showed that a new mineralized bone formed. The histological evaluation of the same case showed that spaces between root fragments of the tooth and bone are filled with connective tissue and the new bone fits tightly with both cementum and dentin [70]. Autogenous teeth grafted as particles were gradually reabsorbed and replaced by new vital bone thanks to osteoinduction and osteoconduction, leading to good implant primary stability. Histological analysis demonstrated that more than 64% of the surface of the implant is in direct contact with bone [78]. There was no difficulty with implant osseointegration thanks to the lack of residual dentin particles that were never in contact with the implant surface. The healing process of the autogenous tooth-derived graft was well harmonized with implant osseointegration demonstrated by a continuous dense mineralized zone of two to five cell layers with osteoblasts that surround the implant surface [78].

Minetti et al. evaluated the histo-morphometric results (new bone in upper jaw sites $37.9 \pm 21.9\%$ and new bone in lower jaw sites $38.0 \pm 22.0\%$) of the application of autogenous tooth-derived bone substitute material in patients treated with alveolus preservation. A total of 101 histological specimens were analyzed by evaluating the total amount of bone, residual dental graft material, and viable bone. The authors stated that alveolar socket preservation employing demineralized autologous tooth-derived biomaterial was a reliable method to obtain fresh vital bone to sustain dental implant rehabilitation [76].

In the study of Sang-Ho Jun et al., tooth bone graft (size 500–1000 μm) and inorganic bovine bone (Bio-Oss) were used as sinus bone grafts in patients with residual bone height less than 500 μm in the maxillary posterior area. After four months the healing state of bone graft sites was evaluated. In a microcomputed tomography analysis, there was no difference in bone density and height between the two groups, instead, in the histo-morphometric analysis, there was a significant difference in the osteoid thickness of the two groups [66].

DDM (size 500–1000 μm) was compared with Bio-Oss as a bone substitute in Guided Bone Regeneration (GBR) in two groups of patients with periodontal teeth and need of implant rehabilitation. GBR was performed with the use of collagen membrane and contextual implant placement. The implant stability, evaluated by Osstell Mentor, and marginal bone resorption, evaluated with x-ray examinations, were measured at T0, at 6 and 18 months after surgery. The values of these parameters were superimposable in the two groups [69].

Minetti et al. studied the use of DDM (406 μm and 815 μm with peaks up to 1110 μm) as a bone substitute for alveolar ridge augmentation with and without collagen membrane. At four months, a bone tissue biopsy to perform and histological analysis showed more bone volume and vital bone in the sites where membrane had been used in association with DDM [79].

DDM can also be used in association with a collagen membrane in post-extraction site preservation and it can be produced from vital or endodontically treated teeth. All filling materials (gutta-percha, cement, etc.) were removed from the teeth. After four months, bone biopsies were performed. The histological and histo-morphometric analyses showed that there was no significant difference in the bone regeneration of sites where DDM was obtained from vital or endodontically treated teeth [37].

Preservation of the post-extraction site of the lower third molar, with tooth bone graft (300–1200 μm) included, ensured bone regeneration of the site and reduced the periodontal defect on the distal side of the lower second molar, which can be created at the site that is left to heal spontaneously [73].

A retrospective study conducted by Korsch and Peichl analyzed the use of autologous dentin for lateral ridge defect reconstruction (tooth-shell technique). This procedure was compared to the bone-shell technique on autogenous bone, according to Khoury. In both groups, implants were placed simultaneously, follow-up was performed at three-month intervals and prosthetic restoration after five months [74]. No significant differences were found between the two groups in terms of complications and outcomes. The integrity of the buccal lamella was preserved in all implants, and all implants had fully osseointegrated, suggesting the use of autologous dentin as a valid and less invasive alternative to autologous bone [74].

In another study, demineralized human dentin and implants were simultaneously implanted in the region of bone defects. No intraoperative or postoperative complications occurred, and no bone resorption was observed. There were no implant or abutment failures in the subsequent 35 months [65].

Kim et al., in a case series, suggested the use of an autogenous fresh demineralized tooth prepared at chairside immediately after extraction for socket preservation. They found that socket preservation using powder, chip, or block type Auto-FDT with a stress-shielding barrier membrane was successful to retain the ridge heights and widths needed for implants [67].

From the data emerging from the histological examinations of Minetti et al., the average vital bone newly formed following the regeneration process using tooth-derived grafts was approximately 27%. This value was significant. A very high bone volume can mean bone containing only graft material that cannot guarantee osteointegration of implants. On the other hand, a high percentage of vital bone indicated a large rate of truly newly formed regenerated vital bone that will participate in the metabolism dynamics and therefore will allow stability and integration [5].

Okubo et al. presented a case of alveolar bone preservation using autogenous primary dental material, with clinical and histological results. The main advantages of the presented technique were great availability in terms of volume, the use of fully autogenous material to reduce the possibility of adverse immune reactions in case of refusal of the patient to receive biomaterials of animal origin, the use of deciduous tooth is completely free of any biological or economic cost, and deciduous teeth, having less enamel than permanent teeth, have greater osteoinduction [77].

In 2015 Park et al. conducted an *in vivo* study on the use of deciduous teeth reduced to powder and used as filler material after appropriate demineralization, concluding that deciduous teeth had adequate structural and physicochemical characteristics suitable to be used as grafting materials [83].

In 2018 Bono et al. described that deciduous teeth could be used as grafting materials in bone augmentation treatments. They also shed light that collagen and BMP-2 protein contents in demineralized tooth material were conserved after chemical treatment. Furthermore, they assessed *in vitro* the response of osteoblastic cells to exogenous stimulation of BMP-2 (at different protein concentrations) to detect the minimum concentration of BMP-2 able to induce the expression of alkaline phosphatase, the marker of early osteoblastic phenotype [68].

A case report, with two-year follow-up, exposed that socket preservation was possible using dental grafting material, and the same site was subsequently treated with implant-supported rehabilitation [72].

Bone remodeling is a constant and complex process for bone tissues in the body and could cause resorption of alveolar bone when a tooth is no longer in place. To compensate for insufficient available bone volume, a number of surgical procedures designed to augment bone volume are described and validated in the scientific literature [72]. As we know from the literature, after tooth extraction, healing of the socket happens through the sequence of many steps, beginning with the stabilization of the clot, the formation of fibrin, and, finally, the recruitment of osteoblasts that will be in charge of the formation of new bone. Several biomaterials and techniques for alveolar volume preservation have been reported, with many functionalities (osteoconduction, osteoinduction, or even stimulation) according to the features of each material [72].

In several clinical situations, autogenous bone grafting has been recognized as the most successful biomaterial. Human dentin and bone exhibit many similarities in regard to mineralization. In fact, after demineralization, both DDM and DBM are made up predominantly of type I collagen (90%) and NCP and, among them, growth factors; therefore, human DDM is a bio-collagen scaffold containing osteoinductive growth factors that supply an appropriate setting to promote new bone formation [72].

According to the study of Cervera-Maillo et al., on 10 partially edentulous patients, a particle-sized dentin graft was performed as an alternative material for socket preservation, split technique, and maxillary sinus elevation. After 24 months of evaluation there was a high bone resorption rate and bone replacement without inflammation, suggesting this as an acceptable biomaterial for several bone defects due to its osteoinductive and osteoconductive properties [75].

Open tubes of dentine particles allowed capillaries to enter, favouring rapid resorption. Clinically and histologically, dentin graft performance is at least comparable to xenogeneic or allogeneic biomaterials [75].

The study of Minetti et al. evaluated the use of the extracted tooth as an autologous graft for socket preservation on patients with post-extraction bone defects. For this purpose, The TT[®] shredding and decontamination machine was used. The graft thus obtained was subsequently inserted at the time of extraction [38].

An innovative preparation method was used, using the dedicated automated TT[®] device, capable of transforming autologous teeth into suitable grafting material. The extracted tooth was cleaned and treated with a dental transformer and socket preservation was performed. Thirteen biopsies were performed to analyze the histological findings over an average time of four months to evaluate if the autologous tooth graft obtained from the root after endodontic therapy should be used in human bone regeneration as a graft for the placement of dental implants [38].

The study of Kadkhodazadeh et al. aimed to evaluate the osteopromoting ability of human tooth powder and compare it with a bovine xenograft, a synthetic material, and a demineralized freeze-dried bone allograft (DFDBA). Tooth powder was able to increase

cell proliferation compared with bovine xenograft, synthetic graft, and DFDBA. However, its osteopromotional capacity was inferior to that of osteogenic materials [71].

The quantity of grafting needed in cases of severe bone deficiencies is a source of discussion. Depending on the tooth, the mean weight and volume obtained by grinding the tooth with TT[®] varied between 0.68 g and 1.88 g and 0.38 cc and 0.96 cc, respectively [5].

In a critical size defect in rats, dual administration of the angiogenic growth factors VEGF and osteogenic BMP-2 demonstrated a remarkable capacity for almost full regeneration [84]. Different studies that evaluated the socket preservation technique with dentin particulate autograft and platelet-rich fibrin concluded that this technique and biomaterial combination had produced well-maintained vertical socket dimensions and minimal horizontal ridge reduction [85,86]. Platelet-rich fibrin is bioactive molecules that can increase cell proliferation and bone remodelling [87,88]. The fibrin matrix's architecture affects the trapping/release of GF [89] Within 7–14 days, platelets and macrophages release a variety of growth factors, such as vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), transforming growth factor- β 1 (TGF- β 1), and insulin-like growth factor (IGF) [89–93].

When there are not enough teeth, Umebayashi et al. recommend using autologous bone or the combination of autologous tooth and other biomaterials [39]. In their study, the autologous tooth matrix was partially demineralised and combined with cancellous and medullary bone particles for bilateral sinus lift and anterior maxillary reconstruction. This combination showed osteoinductive and osteoconductive capabilities in bone regeneration [39].

In another study, partially demineralised dental matrix was combined with xenograft material (Bio-oss) obtaining good results in bone regeneration, even if bovine bone resorption was slower than with the dental matrix [40]. In extensive bone deficiencies, the dental matrix can be coupled with other biomaterials to produce excellent results. [39–42].

5. Devices for Tooth Processing

The material of the tooth can be considered an ideal scaffold due to its osteoconductive properties, which come from its construction with HA and collagen types 1 and 3, and its natural protein content also makes it a material with osteoinductive properties [4,15]. However, this property depends on the amount of proteins that will be found in the material after treatment; therefore, the way of processing this material is very important and still under discussion. Finding a quick, easy procedure that can demineralize, cleanse, and crush teeth while preserving all of their natural properties is the real challenge [5].

In the market to date, there are two devices suitable for using the tooth as a grafting material that have medical CE markings: TT[®] and Bon Maker[®] (Figure 3) [68,94]. Two other devices, Kometabio and Vacuasonic, do not have the medical CE Marking and therefore have not been considered.

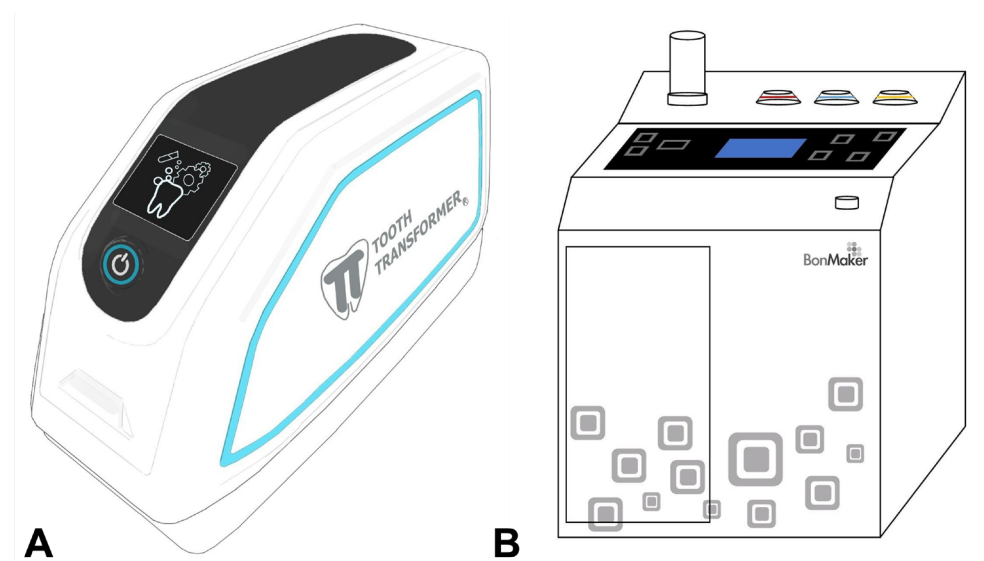


Figure 3. (A) The Tooth Transformer® (CE); (B) BonMaker® (CE).

5.1. Step 1: Tooth Cleaning

After extraction, caries, tartar, soft tissue debris, fillings, cements, and prosthetic parts must be removed from the tooth (Figure 4A). Teeth with root canal therapies can also be used. For the Bon Maker® device, which uses a sieve to separate granules according to size, it is advisable to remove with tweezers the parts that are not consistent with the tooth tissue after grinding. For the TT®, which requires sectioning of the tooth for the trituration step (Figure 4B), cleaning from residual root canal care can be performed during the sectioning stage, simplifying the cleaning procedure for small sections that are more easily visible using magnifiers [5].

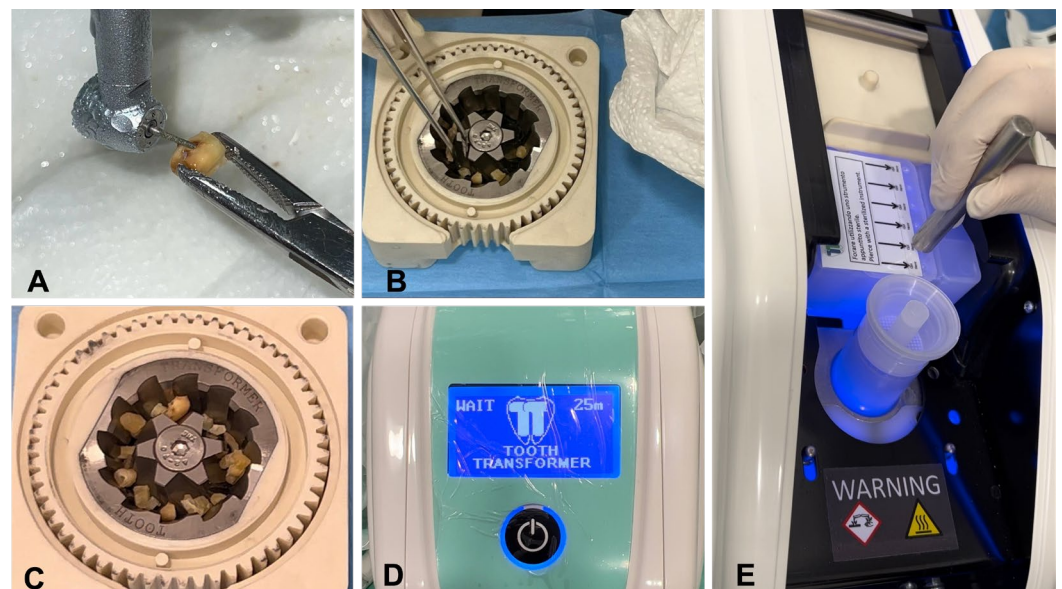


Figure 4. (A) Extracted teeth are cleaned with the use of a diamond bur mounted on a turbine to remove any carious process residue from the tooth surface; (B) Dental elements prepared for grinding; (C) Grinding with tooth fragments positioned in Tooth Grinder® ready for further shredding; (D) Inserted the liquids and pierced the cartridge before the beginning of the transformation cycle; (E) Starting the shredding process with tooth grinding. The transformation cycle lasts only 25 min.

5.2. Step 2: Tooth Grinding

The Bon Maker® device uses a hammer and pestle to crush the tooth, a risky procedure for the operator. Having performed the crushing, the tooth is placed in a non-sterilizable high-speed mill and the granules are separated by a manual sieve, which separates fragments of different sizes by making use of two nets of different filtrations: the larger granules, of 850 µm, are blocked by the first filter, while the finer granules, of 450 µm pass, through the second filter, into the lower plate [5,94]. The TT®, although it has the disadvantage of not being able to insert a whole tooth into the shredder, is equipped with a multi-purpose sterilizable system that works at low speed, which makes it possible to avoid the loss of tooth substance in pulverization [5] (Figure 4C).

5.3. Step 3: Treatment by Device

5.3.1. Bon Maker®

The granules should be manually placed in a sterilizable plastic cylindrical container (Bonbin), which should be inserted into a slot in the machine's upper front. The liquids, which are contained in disposable flasks, must be manually emptied into their respective cavities according to a color code. A bottle to be filled with saline solution and screwed onto the device's top is also required. The material is extracted from the Bonbin at the end of the treatment, which takes about 26 min. The spent and contaminated liquids are collected in a flask glass that is placed behind a door at the front of the device and must be emptied after a few uses. The liquids' composition was analyzed at the Politecnico di Milano, with the following results: HCl 0.45 M-H₂O₂ 130 volumes-ethanol 62.6% chloroform 31.3% water 6.1% + saline wash solution; HCl 0.56 M-H₂O₂ 120 volumes-ethanol 47.2% chloroform 47.2% water 5.6% + saline wash solution [5,95].

5.3.2. Tooth Transformer®

After inserting the tooth into the shredder, the device is closed and inserted. A liquid cartridge and a cylinder with a granulate collection cup (maker) are inserted into the device in their respective housings, the cartridge is activated by punching, and the process begins when the door is closed and the button is pressed (Figure 4D). The liquids are three distinct solutions contained in six distinct compartments of the same single-use cartridge. Two of them are active liquids made up of 0.1 m hydrochloric acid and 10% hydrogen peroxide, while the other is demineralized water. The four compartments filled with mineralized water are employed to eliminate the acid residues in four separate method steps. Following the automatic perforation of the lower cartridge membrane, the six liquids in the cartridge fall by gravity and initiate the process. The procedure is completely automated and repeats the same steps every time. The transformation cycle lasts only 25 min (Figure 4E). The granules are dropped into the collection basket during the first phase of low-speed shredding. To avoid protein damage, the granules are subjected to UVA rays and ultrasonic vibrations with temperature changes, always below 45 °C. At the end of the process, the used and contaminated liquids remain inside their container and can be removed, as they are disposable. [68,96–98].

6. Tooth Transformer® Technique

TT® technology allows for obtaining a demineralized and disinfected tooth material with a pasty consistency [5,94].

The processing of the tooth material with TT® makes it possible to obtain a 400–800 µm graft at the end of the procedure [5,99] (Figure 5). Different studies have been conducted on the size of the graft particles and the impact they have on bone formation. It is said that the graft with larger particle size favors the growth of blood vessels as it leaves more space [100]. Dozza et al., in his study with three different sizes of DBM particles, suggested using an average size of 0.5 to 1 mm for a better result [34]. Koga et al. performed a comparative study between DDM, partial DDM, and mineralized dentin matrix

using three different sizes of graft particles in each group [35]. According to this study, partial demineralization of the dental matrix with particles between 500 and 1000 μm has a larger potential for regeneration than total demineralization since it retains more growth factors that could influence osteogenesis [34,35]. The demineralized dental matrix resorbs faster than mineralized dentin since the biodegradation of big particles with a high crystalline content is quite impossible. The best osteoconduction results come from small-sized HA crystals; however, very small parts could lead to rapid resorption of the graft and failure to preserve the volume [5,36,37,101]. TT[®] has two different speed grinders, the low speed and the high speed. This grinding procedure offers a particle size of the graft between 400 and 800 μm [5]. Scanning Electron Microscope (SEM) analysis revealed that the autologous dental graft's density, roughness, and homogeneity are roughly equivalent to those of autogenous cortical bone, with a surface that contains both mineralized material and organic parts. [36]. The tooth matrix does not lose bone volume over time like other autologous grafts do [101].

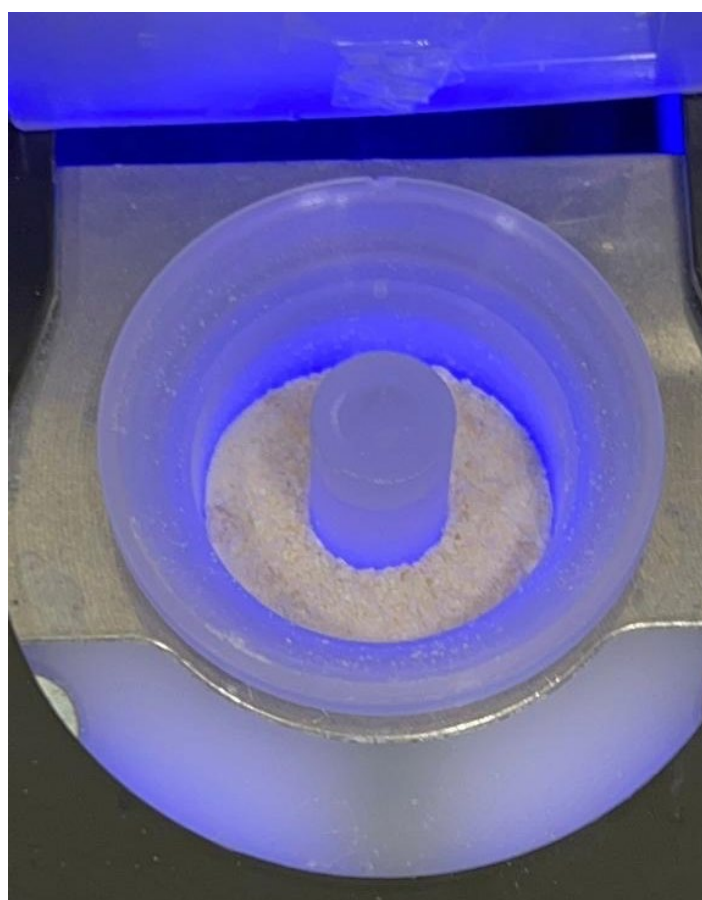


Figure 5. Term processing of the teeth that are shredded between 400 and 1000 μm . Ready for grafting in the oral cavity.

The porosity has an impact on the graft's early fibro-vascularization and new bone replacement. Additionally, the scaffold's pore size has the power to influence how osteogenesis develops. Bigger scaffold pores enable vascularization, which directly signals the start of osteogenesis [102,103]. Dentin is formed by microtubules that are created as a result of the passage of nerve extensions. These tubes have a diameter that varies from 2.5 μm to 0.9 μm with an average of 1.2 μm [104]. The macrophages, mesenchymal cells, and osteoclasts must increase to determine collagenolytic enzyme production so that the resorption process induced by the biomaterial graft begins [105]. Mesenchymal cells, osteoblasts, and other bone cells range in size from 10 to 20 micrometers, while osteoclasts range in size from 20 to 100 μm or more [106,107]. Cell invasion into the dentinal tubules

is not conceivable and the demineralization, by increasing the size of the dentinal tubules, favors the adhesion and the activity of the osteoblasts and their number. Resorption of undecalcified dentin is incomplete or delayed [108]. Minerals seem to make the matrix inaccessible to the action of collagenolytic enzymes. Under the microscope, non-demineralized dentin appears with tubules blocked by mineralized tissue material [109]. Bertassoni et al.'s findings demonstrated that the dentinal peritubular is made up of a collagen-free organic network that is embedded with HA. After acid treatment, the mineral was primarily dissolved by demineralization in peritubular dentin, which had expanded the tubules as well as a meshwork that protruded toward the lumen of the tubules was seen. After demineralization, a dense network of collagen fibrils with amorphous molecules located in it was distinguished [110]. For this reason, undecalcified dentin was not resorbed until 8–12 weeks, much later than decalcified dentin. A notable element is that after one, two, and three weeks of demineralization, the tubule widths in the wet state—which were $1.3 \pm 0.2 \mu\text{m}$ in sound dentine—are $2.5 \pm 0.3 \mu\text{m}$, $2.2 \pm 0.3 \mu\text{m}$, and $1.7 \pm 0.2 \mu\text{m}$, respectively [111]. However, a varied image of the surface with a higher or lesser openness of the tubules is obtained depending on the liquid or combination of liquids utilized. It is believed that a biomaterial's submicronic surface structure encourages osteogenesis by controlling the Transforming Growth Factor (TGF) pathway and causing Mesenchymal Stem Cell (MSC) differentiation [112]. Tanoue et al.'s study showed that osteocyte processes and cytoplasm invaded the demineralized dentinal tubules. The degree of invasion was $5 \mu\text{m}$ away from the new DDM bone contact and osteocyte cellular processes penetrated the dentinal tubules, forming bone tissue that filled the DDM surface [81]. Koga et al. cultivated osteoblasts on the surfaces of mineralized dentin and demineralized dentine matrixes, and only the demineralized dentine matrix surface showed evidence of osteoblast adhesion, this was not observed in mineralized dentin [35]. Therefore, mineral traces on the surface, which correspond to high Ca and P values may limit the ability of the cells to adhere. Different levels of demineralization lead to different concentrations of the Ca/P ratio. The ideal Ca/P ratio in human bones is 1.67 [33]. The TT[®] technique also protects the proteins in the tooth matrix while providing a Ca/P ratio (1.70) that is closer to the natural ratio observed in bone [5]. In contrast to the non-demineralized matrix, this partial demineralization of the tooth obtained from TT[®] decreases the concentration of Ca and P while simultaneously increasing the bioavailability of BMP-2 [68,113]. Bono et al. supported the idea of employing demineralized, treated teeth by providing evidence that BMP-2 is more bioavailable after TT[®] demineralization. Six solutions (liquids) are used in the process to demineralize and disinfect the crushed tooth, giving a pasty consistency [68]. Increased wettability and increased hydrophilicity of the surface are two key features of this surface modification. In comparison to hydrophobic surfaces, hydrophilic surfaces have shown enhanced mineralization, local factor generation, and osteoblast maturation. Microtomography affects osteoblast maturation [114,115]. Some studies claimed that demineralization damages the microstructure of dentin and eliminates the organic material found in it [116,117]. The protocol used by TT[®] with six liquids does not damage the dentinal structure, but BMP-2 availability in the matrix is improved [5,68].

High demineralization substances result in graft materials with low osteogenic potential since they reduce or eliminate the BMP-2 and other proteins found in teeth [5]. Such as those in the small integrin-binding ligand N-linked glycoprotein (SIBLING) family [118]. Natural cell-adhesive and Matrix Metalloproteinases (MMP) binding sites that are crucial for viability, proliferation, and differentiation are retained by the dentin matrix proteins [119]. DMP1, also known as AG1 earlier, was first discovered in teeth but was later discovered in bones, where it is mainly expressed by osteocytes. DMP1 is a multifunctional protein that plays roles in the development of odonto- and osteoblasts, phosphate homeostasis, and the biomineralization of bones and dentin [120–125].

The development of mesenchymal cells into osteoblasts is greatly aided by the BMPs, which belong to the TGFbeta superfamily [27]. Recombinant human BMP-2(rhBMP-2) was authorized for use by the Food and Drug Administration (FDA) due to its range of

uses and osteogenic potential. BMP-2 binds to type I and type II serine/threonine kinase receptors on target cells. This process activates the Smad (canonical) and non-Smad (non-canonical) signaling pathways, which in turn activate osteogenic genes like Osterix and RUNX2. This process stimulates MSC differentiation into osteoblasts [28]. Li et al. detected an increase in the activity of Alkaline Phosphatase (ALP) which results in an increase in osteodifferentiation activity by the BMP-2 [29]. After undergoing TT[®] treatment, the concentration of BMP-2 in the dentin matrix is 22ng/mL, which is higher than the minimum concentration of 12.5ng/mL required to cause considerable ALP activity, as described by Blum et al [5,113,126].

The tooth material processed with the TT[®] technique was applied to fill the alveoli to preserve the bone after tooth extraction (socket preservation technique) [5]. According to the authors, the success rate of the implant following bone regeneration was 99.1%, and it took four months for the bone to repair and be prepared for the implant. BMP-2 and collagen are not removed by the solution's demineralization and decontamination processes. [37,38]. By serving as a carrier for the BMP-2 present in the dental structure, demineralized dentin combines the characteristics of the scaffold with those of the BMP-2. However, substances that cause high demineralization produce dental-derived graft materials with low osteogenic potential because they reduce or eliminate the proteins present in the tooth, including BMP-2.

7. Microbiota and Bone Metabolism

Systemic health is the result of the equilibrium between the various components of the gut. The intestinal environment represents 95% of the microorganisms present in the body. The remainder is present in other environments (for example: mouth, nose, skin, and genitals) [43–45].

The main families of bacteria that constitute the gut microbiota are Actinobacteria, Bacteroides, Firmicutes, Proteobacteria, and Verrucomicrobia. The confirmation of the intestine's neuro–endocrine–immune nature over the past 20 years has been one of science's most important discoveries, known as the gut–brain axis [46,127,128].

There has been much research and discussion on the significance of the gut microbiota in the bone regeneration process [61,129–136]:

- The intestinal microbiota can regulate bone density because it increases the solubility of inorganic salts (calcium, phosphate, and magnesium) and the absorption through the intestinal wall [137,138];
- Gut microbiota homeostasis increases calbindin-D9k expression resulting in increased calcium reabsorption [139];
- The intestinal microbiota regulates the synthesis of serotonin and therefore bone metabolism [140];
- The homeostasis of the intestinal microbiota promotes the proliferation of enterocytes, strengthening the absorption of minerals [141];
- The microbiota ensures the barrier effect of the intestinal epithelium, which prevents "leaky gut" by avoiding the increase in inflammatory cytokines which induce activation of osteoclasts and bone destruction [142,143];
- The intestinal microbiota produces SCFA: SCFAs inhibit nuclear factor kappa B (NF-κB) and therefore inflammation. Furthermore, it reduces the expression of TNF Receptor Associated Factor 6 (TRAF6) and Nuclear Factor Of Activated T Cells 1 (NFATc1) by inhibiting the action of osteoclasts [144,145];
- The intestinal microbiota promotes the production of the hormone IGF-1, which induces the differentiation of osteoblasts [146];
- The intestinal microbiota interacts with the immune system by promoting the production of proinflammatory cytokines with an osteoclastogenic action, determining bone resorption [53,54].

The comprehension of the relationship between gut microbiota and bone mass disorders will be made easier with a greater knowledge of how microorganisms' shape and function change [147,148]. It is still unknown how the gut flora alters in people with osteoporosis. Most likely, the immune–inflammatory axis may serve as a crucial link connecting bone metabolism to the gut flora. According to studies, mice grown germ-free (GF) have more bone mass. In the bone and bone marrow of GF animals, the scientists found fewer osteoclasts, osteoclast precursor cells, CD4 (+) cells, and inflammatory cytokines [149]. Additionally, they claimed that once GF mice had their gut flora transplanted, their bone mass returned to normal [133].

Moreover, studies by Bindels et al. (2015) [150], Maekawa and Hajishengallis (2014) [151], and Scholz-Ahrens et al. (2007) [55] have demonstrated that taking specific pre- and probiotics might enhance bone mass. According to research, gut microbiota and some probiotics may control IGF-1, TNF-, and IL-1, altering bone development and formation [56,152].

Additionally, the intestinal system allows vitamins like folic acid and the vitamin B2-12 complex absorption, permits the synthesis of vitamin K, which is crucial for the vitamin D absorption mechanism, and plays a crucial role in the homeostasis of the skeletal system and in the development of osteocytes, osteoblasts, and osteoclasts [58–60]. SCFAs like acetic acid, which are the sources of energy for both the complex cellular structure that makes up the intestinal epithelium and for the ongoing maintenance of the skeletal system, are produced by the intestine microbiome [57,153].

The link that exists between the bone system and dysbiosis is yet to be investigated. It seems that in a state of dysbiosis, bacterial lipopolysaccharide (LPS) endotoxin increases, promoting endotoxemia and osteoclastic activity [61,129]. The osteoblast and other cells express the androgen receptor (AR) and the estrogen receptor (ER). The effect of estrogens and androgens on bone mass is influenced by osteoclast progenitors [154]. Estrogens may have a protective effect on the maintenance of cortical bone mass as a result of ER signaling that is not nuclear-started. The ER in mesenchymal cells may promote bone apposition from the periosteum [155]. Recent studies demonstrated that the activation of the RANK Ligand/System RANK/Osteoprotegerin (OPG) is partially responsible for the adverse effects of estrogens and their deficit in bone remodeling [43]. Mature osteoclasts convert mononuclear precursors depending on how the RANKL and RANK interact. Because estrogen levels are lower in postmenopausal women, osteoblasts express more RANKL at higher rates, which encourages bone resorption [156,157].

The few results present in the literature showed a distinct kind of bacterium: patients with osteopenia showed an increase in the Firmicutes phyla and a decrease in the Bacteroidetes phyla compared to the control group [152]. Furthermore, subjects with a chronic inflammation (subjects affected by Crohn's disease and ulcerative colitis) present a picture of osteoporosis [158]. Hormones responsible for the production and stabilization of bone tissue are inhibited by the chronic overexpression of pro-inflammatory interleukins, causing the development of osteoporosis [152,159]. It is now known that the microbiota plays an important role in many systemic conditions; therefore, intestinal and oral dysbiosis influence osteoporosis and bone loss [160,161]. Probiotics interacting with intestinal and oral microbiota give good results in various conditions and pathologies [47,162–164]. With the use of probiotics for oral administration, the control of intestinal microbiota is a crucial component in the immune system's response [165].

Nowadays literature confirms the correlation between general health and gut microbiome which focuses on T- and Th17-immune cells. Therefore, dysbiosis can affect the immune system necessary for bone balance. Thus, the therapy of excessive bone resorption and bone healing disorders may benefit from gut microbiota homeostasis [53].

8. Conclusions

In many respects, autologous bone grafts for bone regeneration continue to be the gold standard of bone grafting materials. Demineralized dentin can be used as a grafting material, demonstrating high cell compatibility and rapid bone regeneration due to an

optimal balance between resorption and production of newly formed bone. In the alveolus preservation technique, the use of demineralized dentin and platelet-rich fibrin, in combination with biomaterials, allows good vertical preservation of the alveolus with minimal horizontal reduction. Platelet-Rich Growth Factor (PRGF), Platelet-Derived Growth Factor (PDGF), and Endothelial Growth Factor (VEGF) promote neoangiogenesis and morphogenesis when mixed with bone graft substitutes. The proper process of dentin demineralization allows increased bioavailability of BMP-2 with osteoinductive as well as osteoconductive capabilities. The TT® is a device with system repeatability capabilities: sterilization, automatic distribution of fluids and their concentration, degree of demineralization, constant granule size (400–800 µm), wettability and plasticity of granules, and preservation of BMP-2. The intestinal microbiota interacts with RVS due to its anti-inflammatory and antimicrobial properties by modulating the absorption of vitamins (folic acid and vitamins B2–B12). It synthesizes vitamin K, a key carrier of vitamin D. This study highlights some correlation between bone and gut microbiota, although the link between bone metabolism and dysbiosis needs further study. Advances in tissue engineering and future research should produce more meaningful scientific evidence to understand the efficacy of these treatments and spread into daily clinical use and not be limited to scientific research alone.

Author Contributions: Conceptualization, A.D.I., A.M.I., G.D., G.M., F.L., A.S., D.H., G.M.T., F.I., A.M., M.C.F., F.P. and G.P.; methodology, A.D.I., I.R.B., F.P., G.D., V.S., D.H., F.L., F.I., G.P., F.V., A.P. (Assunta Patano), A.P. (Andrea Palermo), A.S., B.R., C.D.P., G.D., G.M.T. and V.C.; software, I.R.B., F.L., E.d.R., S.B., M.C., D.D.V., G.M.T., S.S., A.S., E.M., G.M., A.D.I., B.R. and F.I.; validation, F.I., F.L., A.M.I., I.R.B., G.P., V.S., A.P. (Assunta Patano), A.P. (Andrea Palermo), F.P., A.S., F.V., S.B., C.D.P. and M.C.; formal analysis, A.D.I., F.P., A.P. (Assunta Patano), A.P. (Andrea Palermo), G.M.T., B.R., D.D.V., F.I. and G.D.; resources, A.D.I., A.M.I., G.M., S.S., C.D.P., S.B., F.V., F.P. and V.S. data curation, A.M., E.M., V.C., I.R.B., E.d.R., F.L., F.I., V.S. and G.D.; writing—original draft preparation, A.D.I., A.M.I., S.S., E.d.R., B.R., F.P., D.D.V., F.I. and G.D.; writing—review and editing, A.P. (Assunta Patano), A.P. (Andrea Palermo), M.C.F., V.C., S.S., M.C., G.P., I.R.B. and A.S., visualization, G.P., C.D.P., F.L., A.P. (Assunta Patano) and F.I.; supervision, A.D.I., A.M.I., F.P., G.M., D.D.V., F.I. and G.D.; project administration, M.C.F., D.H., S.S., G.M., G.M.T., F.L., F.I. and G.D. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

ALP	Alkaline Phosphatase
AR	Androgen Receptor
BMPs	Bone Morphogenetic Proteins
BSP	Bone Sialoprotein
DBM	Demineralized Bone Matrix
DDM	Demineralized Dentin Matrix
DFDBA	Demineralized Freeze-Dried Bone Allograft
DMP1	Dentin Matrix Protein 1
DSPP	Dentin Sialophosphoprotein
ER	Estrogen Receptor

FDA	Food and Drug Administration
FGFs	Fibroblast Growth Factors
GBR	Guided Bone Regeneration
GF	Germ-Free
HA	Hydroxyapatite
IGFII	Insulin Growth Factor II
LPS	Lipopolysaccharide
MMP	Matrix Metalloproteinases
MSC	Mesenchymal Stem Cell
NCP	Non-Collagen Proteins
NF-kB	Nuclear Factor kappa B
NFATc1	Nuclear Factor of Activated T Cells 1
OPG	Osteoprotegerin
OPN	Osteopontin
PDGF	Platelet-Derived Growth Factor
PRGF	Platelet Rich in Growth Factor
rhBMP-2	Recombinant Human BMP-2
RSV	Resveratrol
RUNX2	Runt-Related Transcription Factor 2
SCFA	Short-Chain Fatty Acids
SEM	Scanning Electron Microscope
TCP	Tricalcium Beta-Phosphate
TGF	Transforming Growth Factor
TRAF6	TNF Receptor Associated Factor 6
TT®	Tooth Transformer®
VEGF	Vascular Endothelial Growth Factor

References

1. Quintessence Publishing CO., INC. Guided Bone Regeneration in Implant Dentistry. Available online: http://www.quintpub.com/display_detail.php?psku=B2494#.Y7sD6uzMKdY (accessed on 8 January 2023).
2. Amini, A.R.; Laurencin, C.T.; Nukavarapu, S.P. Bone Tissue Engineering: Recent Advances and Challenges. *Crit. Rev. Biomed. Eng.* **2012**, *40*, 363–408. <https://doi.org/10.1615/critrevbiomedeng.v40.i5.10>.
3. Palla, G.; Fischer, D.S.; Regev, A.; Theis, F.J. Spatial components of molecular tissue biology. *Nat. Biotechnol.* **2022**, *40*, 308–318. <https://doi.org/10.1038/s41587-021-01182-1>.
4. Hazbala, D.; Inchingolo, A.D.; Inchingolo, A.M.; Malcangi, G.; Santacroce, L.; Minetti, E.; di Venere, D.; Limongelli, L.; Bordea, I.R.; Scarano, A.; et al. The Effectiveness of Autologous Demineralized Tooth Graft for the Bone Ridge Preservation: A Systematic Review of the Literature. *J. Biol. Regul. Homeost. Agents* **2021**, *35*, 283–294. <https://doi.org/10.23812/21-2supp1-28>.
5. Minetti, E.; Carasco, A.; Carasco, M.; Giacometti, E.; Ho, K.L.H.; Palermo, A.; Savadori, P.; Taschieri, S. *Il Dente Come Materiale Da Innesto*, 1st ed.; EDRA: Wood Buffalo, AB, Canada, 2020; Volume UNICO; ISBN 978-88-214-5353-3.
6. Migliorini, F.; Cuzzo, F.; Torsiello, E.; Spiezia, F.; Oliva, F.; Maffulli, N. Autologous Bone Grafting in Trauma and Orthopaedic Surgery: An Evidence-Based Narrative Review. *J. Clin. Med.* **2021**, *10*, 4347. <https://doi.org/10.3390/jcm10194347>.
7. Wildemann, B.; Kadow-Romacker, A.; Pruss, A.; Haas, N.P.; Schmidmaier, G. Quantification of growth factors in allogenic bone grafts extracted with three different methods. *Cell Tissue Bank.* **2006**, *8*, 107–114. <https://doi.org/10.1007/s10561-006-9021-0>.
8. Oliveira, É.R.; Nie, L.; Podstawczyk, D.; Allahbakhsh, A.; Ratnayake, J.; Brasil, D.L.; Shavandi, A. Advances in Growth Factor Delivery for Bone Tissue Engineering. *Int. J. Mol. Sci.* **2021**, *22*, 903. <https://doi.org/10.3390/ijms22020903>.

9. Elgali, I.; Omar, O.; Dahlin, C.; Thomsen, P. Guided bone regeneration: Materials and biological mechanisms revisited. *Eur. J. Oral Sci.* **2017**, *125*, 315–337. <https://doi.org/10.1111/eos.12364>.
10. Alkaabi, S.; Alsabri, G.; NatsirKalla, D.; Alavi, S.; Mueller, W.; Forouzanfar, T.; Helder, M. A systematic review on regenerative alveolar graft materials in clinical trials: Risk of bias and meta-analysis. *J. Plast. Reconstr. Aesthetic Surg.* **2021**, *75*, 356–365. <https://doi.org/10.1016/j.bjps.2021.08.026>.
11. von Arx, T.; Cochran, D.L.; Hermann, J.S.; Schenk, R.K.; Higginbottom, F.L.; Buser, D. Lateral ridge augmentation and implant placement: An experimental study evaluating implant osseointegration in different augmentation materials in the canine mandible. *Int. J. Oral Maxillofac. Implant.* **2001**, *16*, 343–354.
12. Piattelli, A.; Degidi, M.; di Stefano, D.A.; Rubini, C.; Fioroni, M.; Strocchi, R. Microvessel Density in Alveolar Ridge Regeneration with Autologous and Alloplastic Bone. *Implant. Dent.* **2002**, *11*, 370–375. <https://doi.org/10.1097/00008505-200211040-00017>.
13. Tang, G.; Liu, Z.; Liu, Y.; Yu, J.; Wang, X.; Tan, Z.; Ye, X. Recent Trends in the Development of Bone Regenerative Biomaterials. *Front. Cell Dev. Biol.* **2021**, *9*, 665813. <https://doi.org/10.3389/fcell.2021.665813>.
14. Sieverts, M.; Obata, Y.; Rosenberg, J.L.; Woolley, W.; Parkinson, D.Y.; Barnard, H.S.; Pelt, D.M.; Acevedo, C. Unraveling the effect of collagen damage on bone fracture using in situ synchrotron microtomography with deep learning. *Commun. Mater.* **2022**, *3*, 1–13. <https://doi.org/10.1038/s43246-022-00296-6>.
15. Kim, Y.-K.; Lee, J.; Um, I.-W.; Kim, K.-W.; Murata, M.; Akazawa, T.; Mitsugi, M. Tooth-derived bone graft material. *J. Korean Assoc. Oral Maxillofac. Surg.* **2013**, *39*, 103–111. <https://doi.org/10.5125/jkaoms.2013.39.3.103>.
16. Butler, W.T.; Ritchie, H. The nature and functional significance of dentin extracellular matrix proteins. *Int. J. Dev. Biol.* **1995**, *39*, 169–179.
17. Chen, J.; Shapiro, H.S.; Sodek, J. Developmental expression of bone sialoprotein mRNA in rat mineralized connective tissues. *J. Bone Miner. Res.* **2009**, *7*, 987–997. <https://doi.org/10.1002/jbmr.5650070816>.
18. Zhang, J.; Tu, Q.; Chen, J. Applications of transgenics in studies of bone sialoprotein. *J. Cell. Physiol.* **2009**, *220*, 30–34. <https://doi.org/10.1002/jcp.21768>.
19. Schmidt-Schultz, T.H.; Schultz, M. Intact growth factors are conserved in the extracellular matrix of ancient human bone and teeth: A storehouse for the study of human evolution in health and disease. *Biol. Chem.* **2005**, *386*, 19. <https://doi.org/10.1515/bc.2005.090>.
20. Lohmann, C.H.; Andreacchio, D.; Koster, G.; Cochran, D.L.; Dean, D.D.; Boyan, B.D.; Schwartz, Z. Tissue response and osteoinduction of human bone grafts in vivo. *Arch. Orthop. Trauma Surg.* **2001**, *121*, 583–590. <https://doi.org/10.1007/s004020100291>.
21. Włodarski, K.H.; Szczęsny, G.; Kuzaka, B.; Włodarski, P. Long-term preservation of Bone Morphogenetic Activity in stored demineralized murine incisors. *Pol. Orthop. Traumatol.* **2013**, *78*, 97–100.
22. Dijke, P.T.; Miyazono, K.; Heldin, C.-H. Signaling via hetero-oligomeric complexes of type I and type II serine/threonine kinase receptors. *Curr. Opin. Cell Biol.* **1996**, *8*, 139–145. [https://doi.org/10.1016/s0955-0674\(96\)80058-5](https://doi.org/10.1016/s0955-0674(96)80058-5).
23. Ripamonti, U.; Ramoshebi, L.N.; Matsaba, T.; Tasker, J.; Crooks, J.; Teare, J. Bone Induction by BMPs/OPs and Related Family Members in Primates. *J. Bone Jt. Surg.* **2001**, *83*, S1–116. <https://doi.org/10.2106/00004623-200100002-00006>.
24. Burchardt, H. The biology of bone graft repair. *Clin. Orthop. Relat. Res.* **1983**, *174*, 28–42.
25. Canalis, E.; Pash, J.; Varghese, S. Skeletal Growth Factors. *Crit. Rev. Eukaryot. Gene Expr.* **1993**, *3*, 155–166.
26. Tabatabaei, F.S.; Torshabi, M. Effects of Non-Collagenous Proteins, TGF- β 1, and PDGF-BB on Viability and Proliferation of Dental Pulp Stem Cells. *J. Oral Maxillofac. Res.* **2016**, *7*, e4. <https://doi.org/10.5037/jomr.2016.7104>.
27. Luu, H.H.; Song, W.-X.; Luo, X.; Manning, D.; Luo, J.; Deng, Z.-L.; Sharff, K.A.; Montag, A.G.; Haydon, R.C.; He, T.-C. Distinct roles of bone morphogenetic proteins in osteogenic differentiation of mesenchymal stem cells. *J. Orthop. Res.* **2007**, *25*, 665–677. <https://doi.org/10.1002/jor.20359>.
28. Chen, D.; Zhao, M.; Mundy, G.R. Bone Morphogenetic Proteins. *Growth Factors* **2004**, *22*, 233–241. <https://doi.org/10.1080/08977190412331279890>.
29. Li, Y.; Fu, G.; Gong, Y.; Li, B.; Li, W.; Liu, D.; Yang, X. BMP-2 Promotes Osteogenic Differentiation of Mesenchymal Stem Cells by Enhancing Mitochondrial Activity. *J. Musculoskelet. Neuronal. Interact.* **2022**, *22*, 123–131.
30. Mehwish, N.; Xu, M.; Zaeem, M.; Lee, B.H. Mussel-Inspired Surface Functionalization of Porous Albumin Cryogels Supporting Synergistic Antibacterial/Antioxidant Activity and Bone-Like Apatite Formation. *Gels* **2022**, *8*, 679. <https://doi.org/10.3390/gels8100679>.
31. Kabir, A.; Murata, M.; Kusano, K.; Zakaria, S.M.; Noor, A.H.M.; Khuda, F.; Hossain, I.; Sultana, S.; Saito, T. Radiological Evaluation of Human Dentin Autografts in Bangladesh. *J. Hard Tissue Biol.* **2014**, *23*, 363–370. <https://doi.org/10.2485/jhtb.23.363>.
32. Boerckel, J.D.; Kolambkar, Y.M.; Dupont, K.M.; Uhrig, B.A.; Phelps, E.A.; Stevens, H.Y.; García, A.J.; Guldborg, R.E. Effects of protein dose and delivery system on BMP-mediated bone regeneration. *Biomaterials* **2011**, *32*, 5241–5251. <https://doi.org/10.1016/j.biomaterials.2011.03.063>.
33. Sotiropoulou, P.; Fountos, G.; Martini, N.; Koukou, V.; Michail, C.; Kandarakis, I.; Nikiforidis, G. Bone calcium/phosphorus ratio determination using dual energy X-ray method. *Phys. Med.* **2015**, *31*, 307–313. <https://doi.org/10.1016/j.ejmp.2015.01.019>.
34. Dozza, B.; Lesci, I.G.; Duchi, S.; della Bella, E.; Martini, L.; Salamanna, F.; Falconi, M.; Cinotti, S.; Fini, M.; Lucarelli, E.; et al. When size matters: Differences in demineralized bone matrix particles affect collagen structure, mesenchymal stem cell behavior, and osteogenic potential. *J. Biomed. Mater. Res. Part A* **2017**, *105*, 1019–1033. <https://doi.org/10.1002/jbm.a.35975>.

35. Koga, T.; Minamizato, T.; Kawai, Y.; Miura, K.-I.; Takashi, I.; Nakatani, Y.; Sumita, Y.; Asahina, I. Bone Regeneration Using Dentin Matrix Depends on the Degree of Demineralization and Particle Size. *PLoS ONE* **2016**, *11*, e0147235. <https://doi.org/10.1371/journal.pone.0147235>.
36. Kim, Y.-K.; Kim, S.-G.; Yun, P.-Y.; Yeo, I.-S.; Jin, S.-C.; Oh, J.-S.; Kim, H.-J.; Yu, S.-K.; Lee, S.-Y.; Kim, J.-S.; et al. Autogenous teeth used for bone grafting: A comparison with traditional grafting materials. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol.* **2014**, *117*, e39–e45. <https://doi.org/10.1016/j.oooo.2012.04.018>.
37. Minetti, E.; Giacometti, E.; Gambardella, U.; Contessi, M.; Ballini, A.; Marenzi, G.; Celko, M.; Mastrangelo, F. Alveolar Socket Preservation with Different Autologous Graft Materials: Preliminary Results of a Multicenter Pilot Study in Human. *Materials* **2020**, *13*, 1153. <https://doi.org/10.3390/ma13051153>.
38. Minetti, E.; Palermo, A.; Ferrante, F.; Schmitz, J.H.; Ho, H.K.L.; Hann, S.N.D.; Giacometti, E.; Gambardella, U.; Contessi, M.; Celko, M.; et al. Autologous Tooth Graft after Endodontical Treated Used for Socket Preservation: A Multicenter Clinical Study. *Appl. Sci.* **2019**, *9*, 5396. <https://doi.org/10.3390/app9245396>.
39. Umabayashi, M.; Ohba, S.; Kurogi, T.; Noda, S.; Asahina, I. Full Regeneration of Maxillary Alveolar Bone Using Autogenous Partially Demineralized Dentin Matrix and Particulate Cancellous Bone and Marrow for Implant-Supported Full Arch Rehabilitation. *J. Oral Implant.* **2020**, *46*, 122–127. <https://doi.org/10.1563/aaid-joi-d-19-00315>.
40. Supplement, D.; Minetti, E.; Palermo, A.; Savadori, P.; Barlattani, A.; Franco, R.; Michele, M.; Gianfreda, F.; Bollero, P. Autologous tooth graft: A histological comparison between dentin mixed with xenograft and dentin alone grafts in socket preservation. *J. Biol. Regul. Homeost. Agents* **2019**, *33*, 189–197.
41. Liu, X.; Li, Q.; Wang, F.; Wang, Z. Maxillary sinus floor augmentation and dental implant placement using dentin matrix protein-1 gene-modified bone marrow stromal cells mixed with deproteinized bovine bone: A comparative study in beagles. *Arch. Oral Biol.* **2016**, *64*, 102–108. <https://doi.org/10.1016/j.archoralbio.2016.01.004>.
42. Andrade, C.; Camino, J.; Nally, M.; Quirynen, M.; Martínez, B.; Pinto, N. Combining autologous particulate dentin, L-PRF, and fibrinogen to create a matrix for predictable ridge preservation: A pilot clinical study. *Clin. Oral Investig.* **2019**, *24*, 1151–1160. <https://doi.org/10.1007/s00784-019-02922-z>.
43. Contaldo, M.; Iтро, A.; Lajolo, C.; Gioco, G.; Inchingolo, F.; Serpico, R. Overview on Osteoporosis, Periodontitis and Oral Dysbiosis: The Emerging Role of Oral Microbiota. *Appl. Sci.* **2020**, *10*, 6000. <https://doi.org/10.3390/app10176000>.
44. Baquero, F.; Nombela, C. The microbiome as a human organ. *Clin. Microbiol. Infect.* **2012**, *18*, 2–4. <https://doi.org/10.1111/j.1469-0691.2012.03916.x>.
45. Inchingolo, A.D.; Cazzolla, A.P.; di Cosola, M.; Greco Lucchina, A.; Santacroce, L.; Charitos, I.A.; Topi, S.; Malcangi, G.; Hazballa, D.; Scarano, A.; et al. The integumentary system and its microbiota between health and disease. *J. Biol. Regul. Homeost. Agents* **2021**, *35*, 303–321. <https://doi.org/10.23812/21-2SUPP1-30>.
46. Pei, Z.; Bini, E.J.; Yang, L.; Zhou, M.; Francois, F.; Blaser, M.J. Bacterial biota in the human distal esophagus. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4250–4255. <https://doi.org/10.1073/pnas.0306398101>.
47. Inchingolo, A.D.; Malcangi, G.; Inchingolo, A.M.; Piras, F.; Settanni, V.; Garofoli, G.; Palmieri, G.; Ceci, S.; Patano, A.; de Leonardi, N.; et al. Benefits and Implications of Resveratrol Supplementation on Microbiota Modulations: A Systematic Review of the Literature. *Int. J. Mol. Sci.* **2022**, *23*, 4027. <https://doi.org/10.3390/ijms23074027>.
48. Inchingolo, A.D.; Inchingolo, A.M.; Malcangi, G.; Avantario, P.; Azzollini, D.; Buongiorno, S.; Viapiano, F.; Campanelli, M.; Ciocia, A.M.; de Leonardi, N.; et al. Effects of Resveratrol, Curcumin and Quercetin Supplementation on Bone Metabolism—A Systematic Review. *Nutrients* **2022**, *14*, 3519. <https://doi.org/10.3390/nu14173519>.
49. Laparra, J.M.; Sanz, Y. Interactions of gut microbiota with functional food components and nutraceuticals. *Pharmacol. Res.* **2010**, *61*, 219–225.
50. Carrera-Quintanar, L.; Roa, R.I.L.; Quintero-Fabián, S.; Sánchez-Sánchez, M.A.; Vizmanos, B.; Ortuño-Sahagún, D. Phytochemicals That Influence Gut Microbiota as Prophyllactics and for the Treatment of Obesity and Inflammatory Diseases. *Mediat. Inflamm.* **2018**, *2018*, 9734845. <https://doi.org/10.1155/2018/9734845>.
51. Bode, L.M.; Bunzel, D.; Huch, M.; Cho, G.-S.; Ruhland, D.; Bunzel, M.; Bub, A.; Franz, C.M.; E Kulling, S. In vivo and in vitro metabolism of trans-resveratrol by human gut microbiota. *Am. J. Clin. Nutr.* **2013**, *97*, 295–309. <https://doi.org/10.3945/ajcn.112.049379>.
52. Nargeh, H.; Aliabadi, F.; Ajami, M.; Pazoki-Toroudi, H. Role of Polyphenols on Gut Microbiota and the Ubiquitin-Proteasome System in Neurodegenerative Diseases. *J. Agric. Food Chem.* **2021**, *69*, 6119–6144. <https://doi.org/10.1021/acs.jafc.1c00923>.
53. Zhang, Y.-W.; Li, Y.-J.; Lu, P.-P.; Dai, G.-C.; Chen, X.-X.; Rui, Y.-F. The modulatory effect and implication of gut microbiota on osteoporosis: From the perspective of “brain–gut–bone” axis. *Food Funct.* **2021**, *12*, 5703–5718. <https://doi.org/10.1039/d0fo03468a>.
54. Yu, M.; Pal, S.; Paterson, C.W.; Li, J.-Y.; Tyagi, A.M.; Adams, J.; Coopersmith, C.M.; Weitzmann, M.N.; Pacifici, R. Ovariectomy induces bone loss via microbial-dependent trafficking of intestinal TNF+ T cells and Th17 cells. *J. Clin. Investig.* **2021**, *131*. <https://doi.org/10.1172/jci143137>.
55. Scholz-Ahrens, K.E.; Ade, P.; Marten, B.; Weber, P.; Timm, W.; Asil, Y.; Glüer, C.-C.; Schrezenmeier, J. Prebiotics, Probiotics, and Synbiotics Affect Mineral Absorption, Bone Mineral Content, and Bone Structure. *J. Nutr.* **2007**, *137*, S838–S846. <https://doi.org/10.1093/jn/137.3.838s>.

56. Ohlsson, C.; Engdahl, C.; Fåk, F.; Andersson, A.; Windahl, S.H.; Farman, H.H.; Movérare-Skrtic, S.; Islander, U.; Sjögren, K. Probiotics Protect Mice from Ovariectomy-Induced Cortical Bone Loss. *PLoS ONE* **2014**, *9*, e92368. <https://doi.org/10.1371/journal.pone.0092368>.
57. Wang, B.; Yao, M.; Lv, L.; Ling, Z.; Li, L. The Human Microbiota in Health and Disease. *Engineering* **2017**, *3*, 71–82. <https://doi.org/10.1016/j.eng.2017.01.008>.
58. Larsbrink, J.; Rogers, T.E.; Hemsworth, G.R.; McKee, L.S.; Tauzin, A.S.; Spadiut, O.; Klintner, S.; Pudlo, N.A.; Urs, K.; Koropatkin, N.M.; et al. A discrete genetic locus confers xyloglucan metabolism in select human gut Bacteroidetes. *Nature* **2014**, *506*, 498–502. <https://doi.org/10.1038/nature12907>.
59. Morowitz, M.J.; Deneff, V.J.; Costello, E.K.; Thomas, B.C.; Poroyko, V.; Relman, D.A.; Banfield, J.F. Strain-resolved community genomic analysis of gut microbial colonization in a premature infant. *Proc. Natl. Acad. Sci. USA* **2010**, *108*, 1128–1133. <https://doi.org/10.1073/pnas.1010992108>.
60. Signorini, L.; Ballini, A.; Arrigoni, R.; de Leonardis, F.; Saini, R.; Cantore, S.; de Vito, D.; Coscia, M.F.; Dipalma, G.; Santacroce, L.; et al. Evaluation of a Nutraceutical Product with Probiotics, Vitamin D, Plus Banaba Leaf Extracts (*Lagerstroemia speciosa*) in Glycemic Control. *Endocr. Metab. Immune Disord.-Drug Targets* **2021**, *21*, 1356–1365. <https://doi.org/10.2174/1871530320666201109115415>.
61. Chen, Y.-C.; Greenbaum, J.; Shen, H.; Deng, H.-W. Association Between Gut Microbiota and Bone Health: Potential Mechanisms and Prospective. *J. Clin. Endocrinol. Metab.* **2017**, *102*, 3635–3646. <https://doi.org/10.1210/jc.2017-00513>.
62. Xu, Z.; Xie, Z.; Sun, J.; Huang, S.; Chen, Y.; Li, C.; Sun, X.; Xia, B.; Tian, L.; Guo, C.; et al. Gut Microbiome Reveals Specific Dysbiosis in Primary Osteoporosis. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 160. <https://doi.org/10.3389/fcimb.2020.00160>.
63. Cairoli, E.; Zhukouskaya, V.V.; Eller-Vainicher, C.; Chiodini, I. Perspectives on osteoporosis therapies. *J. Endocrinol. Investig.* **2015**, *38*, 303–311. <https://doi.org/10.1007/s40618-014-0236-9>.
64. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 Statement: An Updated Guideline for Reporting Systematic Reviews. *BMJ* **2021**, *372*, n71. <https://doi.org/10.1136/bmj.n71>.
65. Tazaki, J.; Murata, M.; Nakanishi, Y.; Ochi, M.; Hirose, Y.; Akazawa, T.; Yodogawa, S.; Hino, J.; Ito, K.; Kitajo, H.; et al. Simultaneous Implantation of Dental Implants and Autogenous Human Dentin. *Key Eng. Mater.* **2011**, *493–494*, 426–429. <https://doi.org/10.4028/www.scientific.net/kem.493-494.426>.
66. Jun, S.-H.; Ahn, J.-S.; Lee, J.-I.; Ahn, K.-J.; Yun, P.-Y.; Kim, Y.-K. A prospective study on the effectiveness of newly developed autogenous tooth bone graft material for sinus bone graft procedure. *J. Adv. Prosthodont.* **2014**, *6*, 528–538. <https://doi.org/10.4047/jap.2014.6.6.528>.
67. Kim, E.-S.; Lee, I.-K.; Kang, J.-Y.; Lee, E.-Y. Various autogenous fresh demineralized tooth forms for alveolar socket preservation in anterior tooth extraction sites: A series of 4 cases. *Maxillofac. Plast. Reconstr. Surg.* **2015**, *37*, 27. <https://doi.org/10.1186/s40902-015-0026-0>.
68. Bono, N.; Tarsini, P.; Candiani, G. Demineralized Dentin and Enamel Matrices as Suitable Substrates for Bone Regeneration. *J. Appl. Biomater. Funct. Mater.* **2017**, *15*, 236–243. <https://doi.org/10.5301/jabfm.5000373>.
69. Li, P.; Zhu, H.; Huang, D. Autogenous DDM versus Bio-Oss granules in GBR for immediate implantation in periodontal post-extraction sites: A prospective clinical study. *Clin. Implant. Dent. Relat. Res.* **2018**, *20*, 923–928. <https://doi.org/10.1111/cid.12667>.
70. Cardaropoli, D.; Nevins, M.; Schupbach, P. New Bone Formation Using an Extracted Tooth as a Biomaterial: A Case Report with Histologic Evidence. *Int. J. Periodontics Restor. Dent.* **2019**, *39*, 157–163. <https://doi.org/10.11607/prd.4045>.
71. Kadkhodazadeh, M.; Fathiazar, A.; Yadegari, Z.; Amid, R. Comparison of osteopromoting ability of human tooth powder with the demineralized freeze-dried bone allograft, a bovine xenograft, and a synthetic graft: An in vitro study. *J. Adv. Periodontol. Implant. Dent.* **2020**, *12*, 19–23. <https://doi.org/10.34172/japid.2020.005>.
72. Minetti, E.; Taschieri, S.; Corbella, S. Autologous Deciduous Tooth-Derived Material for Alveolar Ridge Preservation: A Clinical and Histological Case Report. *Case Rep. Dent.* **2020**, *2020*, 2936878. <https://doi.org/10.1155/2020/2936878>.
73. Sánchez-Labrador, L.; Martín-Ares, M.; Ortega-Aranegui, R.; López-Quiles, J.; Martínez-González, J.M. Autogenous Dentin Graft in Bone Defects after Lower Third Molar Extraction: A Split-Mouth Clinical Trial. *Materials* **2020**, *13*, 3090. <https://doi.org/10.3390/ma13143090>.
74. Korsch, M.; Peichl, M. Retrospective Study: Lateral Ridge Augmentation Using Autogenous Dentin: Tooth-Shell Technique vs. Bone-Shell Technique. *Int. J. Environ. Res. Public Health* **2021**, *18*, 3174. <https://doi.org/10.3390/ijerph18063174>.
75. Cervera-Maillo, J.M.; Morales-Schwarz, D.; Morales-Melendez, H.; Mahesh, L.; Calvo-Guirado, J.L. Autologous Tooth Dentin Graft: A Retrospective Study in Humans. *Medicina* **2021**, *58*, 56. <https://doi.org/10.3390/medicina58010056>.
76. Minetti, E.; Corbella, S.; Taschieri, S.; Canullo, L. Tooth as graft material: Histologic study. *Clin. Implant. Dent. Relat. Res.* **2022**, *24*, 488–496. <https://doi.org/10.1111/cid.13097>.
77. Okubo, N.; Ishikawa, M.; Shakya, M.; Hosono, H.; Maehara, O.; Ohkawara, T.; Ohnishi, S.; Akazawa, T.; Murata, M. Autograft of Demineralized Dentin Matrix Prepared Immediately after Extraction for Horizontal Bone Augmentation of the Anterior Atrophic Maxilla: A First Case of Non-Vital Tooth-Derived Dentin. *J. Hard Tissue Biol.* **2022**, *31*, 47–54. <https://doi.org/10.2485/jhtb.31.47>.

78. Pohl, S.; Prasad, H.; Prasad, S.; Kotsakis, G. Human Histologic Analysis of Implant Osseointegration in a Healed Site Grafted with Nondemineralized Autologous Tooth-Derived Graft Material. *Int. J. Periodontics Restor. Dent.* **2022**, *42*, e199–e207. <https://doi.org/10.11607/prd.4923>.
79. Minetti, E.; Gianfreda, F.; Palermo, A.; Bollero, P. Autogenous Dentin Particulate Graft for Alveolar Ridge Augmentation with and without Use of Collagen Membrane: Preliminary Histological Analysis on Humans. *Materials* **2022**, *15*, 4319. <https://doi.org/10.3390/ma15124319>.
80. Bhaskar, S.N.; Orban, B.J. *Orban's Oral Histology and Embryology*; 9th ed.; Mosby: St. Louis, MO, USA, 1980; ISBN 978-0-8016-4609-6.
81. Tanoue, R.; Ohta, K.; Miyazono, Y.; Iwanaga, J.; Koba, A.; Natori, T.; Iwamoto, O.; Nakamura, K.-I.; Kusakawa, J. Three-dimensional ultrastructural analysis of the interface between an implanted demineralised dentin matrix and the surrounding newly formed bone. *Sci. Rep.* **2018**, *8*, 2858. <https://doi.org/10.1038/s41598-018-21291-3>.
82. Alvares, K. The role of acidic phosphoproteins in biomineralization. *Connect. Tissue Res.* **2014**, *55*, 34–40. <https://doi.org/10.3109/03008207.2013.867336>.
83. Park, M.; Mah, Y.-J.; Kim, D.-H.; Kim, E.-S.; Park, E.-J. Demineralized deciduous tooth as a source of bone graft material: Its biological and physicochemical characteristics. *Oral Surgery Oral Med. Oral Pathol. Oral Radiol.* **2015**, *120*, 307–314. <https://doi.org/10.1016/j.oooo.2015.05.021>.
84. Patel, Z.S.; Young, S.; Tabata, Y.; Jansen, J.A.; Wong, M.E.; Mikos, A.G. Dual delivery of an angiogenic and an osteogenic growth factor for bone regeneration in a critical size defect model. *Bone* **2008**, *43*, 931–940. <https://doi.org/10.1016/j.bone.2008.06.019>.
85. van Orten, A.; Goetz, W.; Billhan, H. Tooth-Derived Granules in Combination with Platelet-Rich Fibrin (“Sticky Tooth”) in Socket Preservation: A Histological Evaluation. *Dent. J.* **2022**, *10*, 29. <https://doi.org/10.3390/dj10020029>.
86. Pohl, S.; Binderman, I.; Tomac, J. Maintenance of Alveolar Ridge Dimensions Utilizing an Extracted Tooth Dentin Particulate Autograft and Platelet-Rich fibrin: A Retrospective Radiographic Cone-Beam Computed Tomography Study. *Materials* **2020**, *13*, 1083. <https://doi.org/10.3390/ma13051083>.
87. Choukroun, J.; Miron, R.J. Platelet rich fibrin: A second-generation platelet concentrate. In *Platelet Rich Fibrin in Regenerative Dentistry: Biological Background and Clinical Indications*; Miron, R.J., Choukroun, J., Eds.; John Wiley & Sons, Ltd: Oxford, UK, 2017; pp. 1–14; ISBN 978-1-119-40679-2.
88. Fujioka-Kobayashi, M.; Miron, R.J. Biological components of platelet rich fibrin: Growth factor release and cellular activity. In *Platelet Rich Fibrin in Regenerative Dentistry: Biological Background and Clinical Indications*; Miron, R.J., Choukroun, J., Eds.; John Wiley & Sons, Ltd: Oxford, UK, 2017; pp. 15–31; ISBN 978-1-119-40679-2.
89. Dohan Ehrenfest, D.M.; Bielecki, T.; Jimbo, R.; Barbé, G.; Del Corso, M.; Inchingolo, F.; Sammartino, G. Do the Fibrin Architecture and Leukocyte Content Influence the Growth Factor Release of Platelet Concentrates? An Evidence-based Answer Comparing a Pure Platelet-Rich Plasma (P-PRP) Gel and a Leukocyte- and Platelet-Rich Fibrin (L-PRF). *Curr. Pharm. Biotechnol.* **2012**, *13*, 1145–1152. <https://doi.org/10.2174/138920112800624382>.
90. Dohan Ehrenfest, D.M.; del Corso, M.; Diss, A.; Mouhyi, J.; Charrier, J.-B. Three-Dimensional Architecture and Cell Composition of a Choukroun's Platelet-Rich Fibrin Clot and Membrane. *J. Periodontol.* **2010**, *81*, 546–555. <https://doi.org/10.1902/jop.2009.090531>.
91. Simonpieri, A.; del Corso, M.; Vervelle, A.; Jimbo, R.; Inchingolo, F.; Sammartino, G.; Dohan Ehrenfest, D.M. Current knowledge and perspectives for the use of platelet-rich plasma (PRP) and platelet-rich fibrin (PRF) in oral and maxillofacial surgery part 2: Bone graft, implant and reconstructive surgery. *Curr. Pharm. Biotechnol.* **2012**, *13*, 1231–1256. <https://doi.org/10.2174/138920112800624472>.
92. del Corso, M.; Vervelle, A.; Simonpieri, A.; Jimbo, R.; Inchingolo, F.; Sammartino, G.; Dohan Ehrenfest, D.M. Current Knowledge and Perspectives for the Use of Platelet-Rich Plasma (PRP) and Platelet-Rich Fibrin (PRF) in Oral and Maxillofacial Surgery Part 1: Periodontal and Dentoalveolar Surgery. *Curr. Pharm. Biotechnol.* **2012**, *13*, 1207–1230. <https://doi.org/10.2174/138920112800624391>.
93. Dohan Ehrenfest, D.M.; Bielecki, T.; Mishra, A.; Borzini, P.; Inchingolo, F.; Sammartino, G.; Rasmusson, L.; Everts, P.A. In Search of a Consensus Terminology in the Field of Platelet Concentrates for Surgical Use: Platelet-Rich Plasma (PRP), Platelet-Rich Fibrin (PRF), Fibrin Gel Polymerization and Leukocytes. *Curr. Pharm. Biotechnol.* **2012**, *13*, 1131–1137. <https://doi.org/10.2174/138920112800624328>.
94. Kanazirski, N.; Kanazirska, P. Auto-tooth bone graft material for reconstruction of bone defects in the oral region: Case reports. *Folia Medica* **2022**, *64*, 162–168. <https://doi.org/10.3897/folmed.64.e59099>.
95. Kim, K.-W. Bone Induction by Demineralized Dentin Matrix in Nude Mouse Muscles. *Maxillofac. Plast. Reconstr. Surg.* **2014**, *36*, 50–56. <https://doi.org/10.14402/jkamprs.2014.36.2.50>.
96. Eriksson, R.A.; Albrektsson, T. The effect of heat on bone regeneration: An experimental study in the rabbit using the bone growth chamber. *J. Oral Maxillofac. Surg.* **1984**, *42*, 705–711. [https://doi.org/10.1016/0278-2391\(84\)90417-8](https://doi.org/10.1016/0278-2391(84)90417-8).
97. Bischof, J.C.; He, X. Thermal Stability of Proteins. *Ann. N. Y. Acad. Sci.* **2005**, *1066*, 12–33. <https://doi.org/10.1196/annals.1363.003>.
98. Moharamzadeh, K.; Freeman, C.; Blackwood, K. Processed bovine dentine as a bone substitute. *Br. J. Oral Maxillofac. Surg.* **2008**, *46*, 110–113. <https://doi.org/10.1016/j.bjoms.2007.07.209>.

99. Minetti, E.; Berardini, M.; Trisi, P. A New Tooth Processing Apparatus Allowing to Obtain Dentin Grafts for Bone Augmentation: The Tooth Transformer. *Open Dent. J.* **2019**, *13*, 6–14. <https://doi.org/10.2174/1874210601913010006>.
100. Carano, R.A.; Filvaroff, E.H. Angiogenesis and bone repair. *Drug Discov. Today* **2003**, *8*, 980–989. [https://doi.org/10.1016/s1359-6446\(03\)02866-6](https://doi.org/10.1016/s1359-6446(03)02866-6).
101. Minetti, E.; Palermo, A.; Contessi, M.; Gambardella, U.; Schmitz, J.H.; Giacometti, E.; Celko, M.; Trisi, P. Autologous tooth graft for maxillary sinus augmentation: A multicenter clinical study. *Int. J. Growth Factors Stem Cells Dent.* **2019**, *2*, 45. https://doi.org/10.4103/gfsc.gfsc_13_19.
102. Huttmacher, D.W.; Schantz, J.T.; Lam, C.X.F.; Tan, K.C.; Lim, T.C. State of the art and future directions of scaffold-based bone engineering from a biomaterials perspective. *J. Tissue Eng. Regen. Med.* **2007**, *1*, 245–260. <https://doi.org/10.1002/term.24>.
103. Nava, M.M.; Draghi, L.; Giordano, C.; Pietrabissa, R. The Effect of Scaffold Pore Size in Cartilage Tissue Engineering. *J. Appl. Biomater. Funct. Mater.* **2016**, *14*, e223–e229. <https://doi.org/10.5301/jabfm.5000302>.
104. Passos, A.D.; Tziafas, D.; Mouza, A.A.; Paras, S.V. Computational Modelling for Efficient Transdental Drug Delivery. *Fluids* **2017**, *3*, 4. <https://doi.org/10.3390/fluids3010004>.
105. Sheikh, Z.; Abdallah, M.-N.; Hanafi, A.A.; Misbahuddin, S.; Rashid, H.; Glogauer, M. Mechanisms of in Vivo Degradation and Resorption of Calcium Phosphate Based Biomaterials. *Materials* **2015**, *8*, 7913–7925. <https://doi.org/10.3390/ma8115430>.
106. Tiedemann, K.; le Nihouannen, D.; Fong, J.E.; Hussein, O.; Barralet, J.E.; Komarova, S.V. Regulation of Osteoclast Growth and Fusion by mTOR/raptor and mTOR/riCTOR/Akt. *Front. Cell Dev. Biol.* **2017**, *5*, 54. <https://doi.org/10.3389/fcell.2017.00054>.
107. ScienceDirect Topics. Osteoblast—An Overview. Available online: <https://www.sciencedirect.com/topics/materials-science/osteoblast> (accessed on 25 December 2022).
108. Pohl, V.; Pohl, S.; Sulzbacher, I.; Fuerhauser, R.; Mailath-Pokorny, G.; Haas, R. Alveolar Ridge Augmentation Using Dystopic Autogenous Tooth: 2-Year Results of an Open Prospective Study. *Int. J. Oral Maxillofac. Implant.* **2017**, *32*, 870–879. <https://doi.org/10.11607/jomi.5396>.
109. Sohn, D.-S.; Tulio, A.-V.M.; Kang, C.-D.; Fabian, O.-A.; Kim, G.E.A.; Kim, H.-G. Socket preservation using demineralized tooth graft: A case series report with histological analysis. *Int. J. Growth Factors Stem Cells Dent.* **2020**, *3*, 27. https://doi.org/10.4103/gfsc.gfsc_16_19.
110. Bertassoni, L.E.; Stankoska, K.; Swain, M.V. Insights into the structure and composition of the peritubular dentin organic matrix and the lamina limitans. *Micron* **2012**, *43*, 229–236. <https://doi.org/10.1016/j.micron.2011.08.003>.
111. Arends, J.; Stokroos, I.; Jongbloed, W.; Ruben, J. The Diameter of Dentinal Tubules in Human Coronal Dentine after Demineralization and Air Drying. *Caries Res.* **1995**, *29*, 118–121. <https://doi.org/10.1159/000262052>.
112. Miron, R.J.; Zhang, Y. Next-Generation Biomaterials for Bone & Periodontal Regeneration. Available online: http://www.quintpub.com/display_detail.php3?psku=B7963 (accessed on 30 October 2020).
113. Bono, N.; Tarsini, P.; Candiani, G. BMP-2 and type I collagen preservation in human deciduous teeth after demineralization. *J. Appl. Biomater. Funct. Mater.* **2018**, *17*, 2280800018784230. <https://doi.org/10.1177/2280800018784230>.
114. Lim, J.Y.; Taylor, A.F.; Li, Z.; Vogler, E.A.; Donahue, H.J. Integrin Expression and Osteopontin Regulation in Human Fetal Osteoblastic Cells Mediated by Substratum Surface Characteristics. *Tissue Eng.* **2005**, *11*, 19–29. <https://doi.org/10.1089/ten.2005.11.19>.
115. Zhao, G.; Schwartz, Z.; Wieland, M.; Rupp, F.; Geis-Gerstorfer, J.; Cochran, D.L.; Boyan, B.D. High surface energy enhances cell response to titanium substrate microstructure. *J. Biomed. Mater. Res. Part A* **2005**, *74A*, 49–58. <https://doi.org/10.1002/jbm.a.30320>.
116. Kim, Y.-K.; Lee, J.; Kim, K.-W.; Um, I.-W.; Murata, M.; Ito, K. Analysis of Organic Components and Osteoinductivity in Autogenous Tooth Bone Graft Material. *J. Korean Assoc. Maxillofac. Plast. Reconstr. Surg.* **2013**, *35*, 353–359. <https://doi.org/10.14402/jkamprs.2013.35.6.353>.
117. Hedenbjörk-Lager, A.; Hamberg, K.; Pääkkönen, V.; Tjäderhane, L.; Ericson, D. Collagen degradation and preservation of MMP-8 activity in human dentine matrix after demineralization. *Arch. Oral Biol.* **2016**, *68*, 66–72. <https://doi.org/10.1016/j.archoralbio.2016.04.003>.
118. Bertassoni, L.E. Dentin on the nanoscale: Hierarchical organization, mechanical behavior and bioinspired engineering. *Dent. Mater.* **2017**, *33*, 637–649. <https://doi.org/10.1016/j.dental.2017.03.008>.
119. Athirasala, A.; Tahayeri, A.; Thrivikraman, G.; Franca, C.M.; Monteiro, N.; Tran, V.; Ferracane, J.; E Bertassoni, L. A dentin-derived hydrogel bioink for 3D bioprinting of cell laden scaffolds for regenerative dentistry. *Biofabrication* **2017**, *10*, 024101. <https://doi.org/10.1088/1758-5090/aa9b4e>.
120. Toyosawa, S.; Shintani, S.; Fujiwara, T.; Ooshima, T.; Sato, A.; Ijuhin, N.; Komori, T. Dentin Matrix Protein 1 Is Predominantly Expressed in Chicken and Rat Osteocytes but Not in Osteoblasts. *J. Bone Miner. Res.* **2001**, *16*, 2017–2026. <https://doi.org/10.1359/jbmr.2001.16.11.2017>.
121. Ling, Y.; Rios, H.F.; Myers, E.R.; Lu, Y.; Feng, J.Q.; Boskey, A.L. DMP1 depletion decreases bone mineralization in vivo: An FTIR imaging analysis. *J. Bone Miner. Res.* **2005**, *20*, 2169–2177. <https://doi.org/10.1359/JBMR.050815>.
122. Lu, Y.; Ye, L.; Yu, S.; Zhang, S.; Xie, Y.; McKee, M.D.; Li, Y.C.; Kong, J.; Eick, J.D.; Dallas, S.L.; et al. Rescue of odontogenesis in Dmp1-deficient mice by targeted re-expression of DMP1 reveals roles for DMP1 in early odontogenesis and dentin apposition in vivo. *Dev. Biol.* **2007**, *303*, 191–201. <https://doi.org/10.1016/j.ydbio.2006.11.001>.

123. Beniash, E.; Deshpande, A.S.; Fang, P.A.; Lieb, N.S.; Zhang, X.; Sfeir, C.S. Possible role of DMP1 in dentin mineralization. *J. Struct. Biol.* **2011**, *174*, 100–106. <https://doi.org/10.1016/j.jsb.2010.11.013>.
124. Almushayt, A.; Narayanan, K.; E Zaki, A.; George, A. Dentin matrix protein 1 induces cytodifferentiation of dental pulp stem cells into odontoblasts. *Gene Ther.* **2005**, *13*, 611–620. <https://doi.org/10.1038/sj.gt.3302687>.
125. Narayanan, K.; Srinivas, R.; Ramachandran, A.; Hao, J.; Quinn, B.; George, A. Differentiation of embryonic mesenchymal cells to odontoblast-like cells by overexpression of dentin matrix protein 1. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 4516–4521. <https://doi.org/10.1073/pnas.081075198>.
126. Blum, J.S.; Li, R.H.; Mikos, A.G.; Barry, M.A. An optimized method for the chemiluminescent detection of alkaline phosphatase levels during osteodifferentiation by bone morphogenetic protein 2. *J. Cell. Biochem.* **2000**, *80*, 532–537. [https://doi.org/10.1002/1097-4644\(20010315\)80:4<532::aid-jcb1007>3.0.co;2-b](https://doi.org/10.1002/1097-4644(20010315)80:4<532::aid-jcb1007>3.0.co;2-b).
127. Inchingolo, F.; DiPalma, G.; Cirulli, N.; Cantore, S.; Saini, R.S.; Altini, V.; Santacroce, L.; Ballini, A.; Saini, R. Microbiological results of improvement in periodontal condition by administration of oral probiotics. *J. Biol. Regul. Homeost. Agents* **2018**, *32*, 1323–1328.
128. Isacco, C.G.; Ballini, A.; de Vito, D.; Nguyen, K.C.D.; Cantore, S.; Bottalico, L.; Quagliuolo, L.; Boccellino, M.; Di Domenico, M.; Santacroce, L.; et al. Rebalancing the Oral Microbiota as an Efficient Tool in Endocrine, Metabolic and Immune Disorders. *Endocr. Metab. Immune Disord.-Drug Targets* **2021**, *21*, 777–784. <https://doi.org/10.2174/1871530320666200729142504>.
129. Behera, J.; Ison, J.; Tyagi, S.C.; Tyagi, N. The role of gut microbiota in bone homeostasis. *Bone* **2020**, *135*, 115317. <https://doi.org/10.1016/j.bone.2020.115317>.
130. McCabe, L.R.; Britton, R.A.; Parameswaran, N. Prebiotic and Probiotic Regulation of Bone Health: Role of the Intestine and its Microbiome. *Curr. Osteoporos. Rep.* **2015**, *13*, 363–371. <https://doi.org/10.1007/s11914-015-0292-x>.
131. Isacco, C.G.; Ballini, A.; Paduanelli, G.; Inchingolo, A.D.; Nguyen, K.C.D.; Inchingolo, A.M.; Pham, V.H.; Aityan, S.K.; Schiffman, M.; Tran, T.C.; et al. Bone decay and beyond: How can we approach it better. *J. Biol. Regul. Homeost. Agents* **2019**, *33*, 143–154.
132. Das, M.; Cronin, O.; Keohane, D.M.; Cormac, E.M.; Nugent, H.; Nugent, M.; Molloy, C.; O’Toole, P.W.; Shanahan, F.; Molloy, M.G.; et al. Gut microbiota alterations associated with reduced bone mineral density in older adults. *Rheumatology* **2019**, *58*, 2295–2304. <https://doi.org/10.1093/rheumatology/kez302>.
133. Sjögren, K.; Engdahl, C.; Henning, P.; Lerner, U.H.; Tremaroli, V.; Lagerquist, M.K.; Bäckhed, F.; Ohlsson, C. The gut microbiota regulates bone mass in mice. *J. Bone Miner. Res.* **2012**, *27*, 1357–1367. <https://doi.org/10.1002/jbmr.1588>.
134. Weaver, C.M. Diet, Gut Microbiome, and Bone Health. *Curr. Osteoporos. Rep.* **2015**, *13*, 125–130. <https://doi.org/10.1007/s11914-015-0257-0>.
135. Ballini, A.; DiPalma, G.; Isacco, C.G.; Boccellino, M.; Di Di Domenico, M.; Santacroce, L.; Nguyễn, K.C.; Scacco, S.; Calvani, M.; Boddi, A.; et al. Oral Microbiota and Immune System Crosstalk: A Translational Research. *Biology* **2020**, *9*, 131. <https://doi.org/10.3390/biology9060131>.
136. Seely, K.D.; Kotelko, C.A.; Douglas, H.; Bealer, B.; Brooks, A.E. The Human Gut Microbiota: A Key Mediator of Osteoporosis and Osteogenesis. *Int. J. Mol. Sci.* **2021**, *22*, 9452. <https://doi.org/10.3390/ijms22179452>.
137. Bongers, A.; Heuvel, E.G.H.M.V.D. Prebiotics and the Bioavailability of Minerals and Trace Elements. *Food Rev. Int.* **2003**, *19*, 397–422. <https://doi.org/10.1081/fri-120025482>.
138. Qin, L.; Liu, W.; Cao, H.; Xiao, G. Molecular mechanosensors in osteocytes. *Bone Res.* **2020**, *8*, 1–24. <https://doi.org/10.1038/s41413-020-0099-y>.
139. Fukushima, A.; Aizaki, Y.; Sakuma, K. Short-chain fatty acids increase the level of calbindin-D9k messenger RNA in Caco-2 cells. *J. Nutr. Sci. Vitaminol.* **2012**, *58*, 287–291. <https://doi.org/10.3177/jnsv.58.287>.
140. D’Amelio, P.; Sassi, F. Gut Microbiota, Immune System, and Bone. *Calcif. Tissue Int.* **2017**, *102*, 415–425. <https://doi.org/10.1007/s00223-017-0331-y>.
141. Litvak, Y.; Byndloss, M.X.; Bäuml, A.J. Colonocyte metabolism shapes the gut microbiota. *Science* **2018**, *362*, eaat9076. <https://doi.org/10.1126/science.aat9076>.
142. Suzuki, T. Regulation of the intestinal barrier by nutrients: The role of tight junctions. *Anim. Sci. J.* **2020**, *91*, e13357. <https://doi.org/10.1111/asj.13357>.
143. Redlich, K.; Smolen, J.S. Inflammatory bone loss: Pathogenesis and therapeutic intervention. *Nat. Rev. Drug Discov.* **2012**, *11*, 234–250. <https://doi.org/10.1038/nrd3669>.
144. Lucas, S.; Omata, Y.; Hofmann, J.; Böttcher, M.; Iljazovic, A.; Sarter, K.; Albrecht, O.; Schulz, O.; Krishnacoumar, B.; Krönke, G.; et al. Short-chain fatty acids regulate systemic bone mass and protect from pathological bone loss. *Nat. Commun.* **2018**, *9*, 55. <https://doi.org/10.1038/s41467-017-02490-4>.
145. Park, J.-S.; Lee, E.-J.; Lee, J.-C.; Kim, W.-K.; Kim, H.-S. Anti-inflammatory effects of short chain fatty acids in IFN- γ -stimulated RAW 264.7 murine macrophage cells: Involvement of NF- κ B and ERK signaling pathways. *Int. Immunopharmacol.* **2007**, *7*, 70–77. <https://doi.org/10.1016/j.intimp.2006.08.015>.
146. Yan, J.; Herzog, J.W.; Tsang, K.; Brennan, C.A.; Bower, M.A.; Garrett, W.S.; Sartor, B.R.; Aliprantis, A.O.; Charles, J.F. Gut microbiota induce IGF-1 and promote bone formation and growth. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E7554–E7563. <https://doi.org/10.1073/pnas.1607235113>.

147. Jarchum, I.; Pamer, E.G. Regulation of innate and adaptive immunity by the commensal microbiota. *Curr. Opin. Immunol.* **2011**, *23*, 353–360. <https://doi.org/10.1016/j.coi.2011.03.001>.
148. MacPherson, A.J.; Harris, N.L. Interactions between commensal intestinal bacteria and the immune system. *Nat. Rev. Immunol.* **2004**, *4*, 478–485. <https://doi.org/10.1038/nri1373>.
149. Mazmanian, S.K.; Liu, C.H.; Tzianabos, A.O.; Kasper, D.L. An Immunomodulatory Molecule of Symbiotic Bacteria Directs Maturation of the Host Immune System. *Cell* **2005**, *122*, 107–118. <https://doi.org/10.1016/j.cell.2005.05.007>.
150. Bindels, L.B.; Delzenne, N.M.; Cani, P.D.; Walter, J. Towards a more comprehensive concept for prebiotics. *Nat. Rev. Gastroenterol. Hepatol.* **2015**, *12*, 303–310. <https://doi.org/10.1038/nrgastro.2015.47>.
151. Maekawa, T.; Hajishengallis, G. Topical treatment with probiotic *Lactobacillus brevis* CD2 inhibits experimental periodontal inflammation and bone loss. *J. Periodontol. Res.* **2014**, *49*, 785–791. <https://doi.org/10.1111/jre.12164>.
152. Wang, J.; Wang, Y.; Gao, W.; Wang, B.; Zhao, H.; Zeng, Y.; Ji, Y.; Hao, D. Diversity analysis of gut microbiota in osteoporosis and osteopenia patients. *PeerJ* **2017**, *5*, e3450. <https://doi.org/10.7717/peerj.3450>.
153. Campanella, V.; Syed, J.; Santacroce, L.; Saini, R.; Ballini, A.; Inchingolo, F. Oral probiotics influence oral and respiratory tract infections in pediatric population: A randomized double-blinded placebo-controlled pilot study. *Eur. Rev. Med. Pharmacol. Sci.* **2018**, *22*, 8034–8041.
154. Boyce, B.F.; Xing, L. The RANKL/RANK/OPG pathway. *Curr. Osteoporos. Rep.* **2007**, *5*, 98–104. <https://doi.org/10.1007/s11914-007-0024-y>.
155. Park, J.H.; Lee, N.K.; Lee, S.Y. Current Understanding of RANK Signaling in Osteoclast Differentiation and Maturation. *Mol. Cells* **2017**, *40*, 706–713. <https://doi.org/10.14348/molcells.2017.0225>.
156. Walsh, M.C.; Kim, N.; Kadono, Y.; Rho, J.; Lee, S.Y.; Lorenzo, J.; Choi, Y. OSTEOIMMUNOLOGY: Interplay Between the Immune System and Bone Metabolism. *Annu. Rev. Immunol.* **2006**, *24*, 33–63. <https://doi.org/10.1146/annurev.immunol.24.021605.090646>.
157. Borsani, E.; Bonazza, V.; Buffoli, B.; Nocini, P.F.; Albanese, M.; Zotti, F.; Inchingolo, F.; Rezzani, R.; Rodella, L.F. Beneficial Effects of Concentrated Growth Factors and Resveratrol on Human Osteoblasts In Vitro Treated with Bisphosphonates. *BioMed Res. Int.* **2018**, *2018*, 4597321. <https://doi.org/10.1155/2018/4597321>.
158. Inchingolo, F.; Hazballa, D.; Inchingolo, A.D.; Malcangi, G.; Marinelli, G.; Mancini, A.; Maggiore, M.E.; Bordea, I.R.; Scarano, A.; Farronato, M.; et al. Innovative Concepts and Recent Breakthrough for Engineered Graft and Constructs for Bone Regeneration: A Literature Systematic Review. *Materials* **2022**, *15*, 1120. <https://doi.org/10.3390/ma15031120>.
159. Inchingolo, F.; Martelli, F.S.; Gargiulo Isacco, C.; Borsani, E.; Cantore, S.; Corcioli, F.; Boddi, A.; Nguyễn, K.C.D.; De Vito, D.; Aityan, S.K.; et al. Chronic Periodontitis and Immunity, Towards the Implementation of a Personalized Medicine: A Translational Research on Gene Single Nucleotide Polymorphisms (SNPs) Linked to Chronic Oral Dysbiosis in 96 Caucasian Patients. *Biomedicines* **2020**, *8*, 115. <https://doi.org/10.3390/biomedicines8050115>.
160. Isacco, C.G.; Ballini, A.; de Vito, D.; Inchingolo, A.M.; Cantore, S.; Paduanelli, G.; Nguyen, K.C.D.; Inchingolo, A.D.; Dipalma, G.; Inchingolo, F. Probiotics in health and immunity: A first step toward understanding the importance of microbiota system in translational medicine. In *Prebiotics and Probiotics*; IntechOpen: London, UK, 2020. <https://doi.org/10.5772/intechopen.88601>.
161. Cantore, S.; Ballini, A.; de Vito, D.; Abbinante, A.; Altini, V.; DiPalma, G.; Inchingolo, F.; Saini, R. Clinical results of improvement in periodontal condition by administration of oral probiotics. *J. Biol. Regul. Homeost. Agents* **2018**, *32*, 1329–1334.
162. Santacroce, L.; Charitos, I.A.; Ballini, A.; Inchingolo, F.; Luperto, P.; de Nitto, E.; Topi, S. The Human Respiratory System and its Microbiome at a Glimpse. *Biology* **2020**, *9*, 318. <https://doi.org/10.3390/biology9100318>.
163. Inchingolo, F.; Santacroce, L.; Cantore, S.; Ballini, A.; del Prete, R.; Topi, S.; Saini, R.; Dipalma, G.; Arrigoni, R. Probiotics and EpiCor® in human health. *J. Biol. Regul. Homeost. Agents* **2019**, *33*, 1973–1979. <https://doi.org/10.23812/19-543-L>.
164. Ballini, A.; Santacroce, L.; Cantore, S.; Bottalico, L.; Dipalma, G.; Topi, S.; Saini, R.; De Vito, D.; Inchingolo, F. Probiotics Efficacy on Oxidative Stress Values in Inflammatory Bowel Disease: A Randomized Double-Blinded Placebo-Controlled Pilot Study. *Endocr. Metab. Immune Disord.-Drug Targets* **2019**, *19*, 373–381. <https://doi.org/10.2174/1871530319666181221150352>.
165. Inchingolo, A.; Inchingolo, A.; Bordea, I.; Malcangi, G.; Xhajanka, E.; Scarano, A.; Lorusso, F.; Farronato, M.; Tartaglia, G.; Isacco, C.; et al. SARS-CoV-2 Disease through Viral Genomic and Receptor Implications: An Overview of Diagnostic and Immunology Breakthroughs. *Microorganisms* **2021**, *9*, 793. <https://doi.org/10.3390/microorganisms9040793>.

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.